

Role of LDL Cholesterol and Endolysosomes in Amyloidogenesis and Alzheimer's Disease

Xuesong Chen*, Liang Hui, and Jonathan D Geiger

Department of Basic Biomedical Sciences, School of Medicine and Health Sciences, University of North Dakota, Grand Forks, ND 58203, USA

*Corresponding author: Xuesong Chen, Department of Basic Biomedical Sciences, University of North Dakota School of Medicine and Health Sciences 504 Hamline St Grand Forks, North Dakota 58203 USA, Tel: (701) 777-0919; Fax: (701) 777-0387; E-mail: xuesong.chen@med.und.edu

Received date: July 14, 2014, Accepted date: Sep 16, 2014, Published date: Sep 22, 2014

Copyright: © 2014 Chen X, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Abstract

The pathogenesis of late-onset sporadic Alzheimer's disease (AD) is believed to result from complex interactions between nutritional, environmental, epigenetic and genetic factors. Among those factors, altered circulating cholesterol homeostasis, independent of the APOE genotype, continues to be implicated in brain deposition of amyloid beta protein (A β) and the pathogenesis of AD. It is believed that trafficking of amyloid beta precursor protein (A β PP) into endolysosomes appears to play a critical role in determining amyloidogenic processing of A β PP because this is precisely where two enzymes critically important in A β PP metabolism are located; beta amyloid converting enzyme (BACE-1) and gamma secretase enzyme. We have shown that elevated levels of LDL cholesterol promote A β PP internalization, disturb neuronal endolysosome structure and function, and increase A β accumulation in neuronal endolysosomes. Here, we will further discuss the linkage between elevated levels of LDL cholesterol and AD pathogenesis, and explore the underlying mechanisms whereby elevated levels of plasma LDL cholesterol promote amyloidogenesis.

Keywords: LDL cholesterol; Amyloid beta; Amyloid precursor protein; Sporadic Alzheimer's disease; Endosome; Lysosome

Introduction

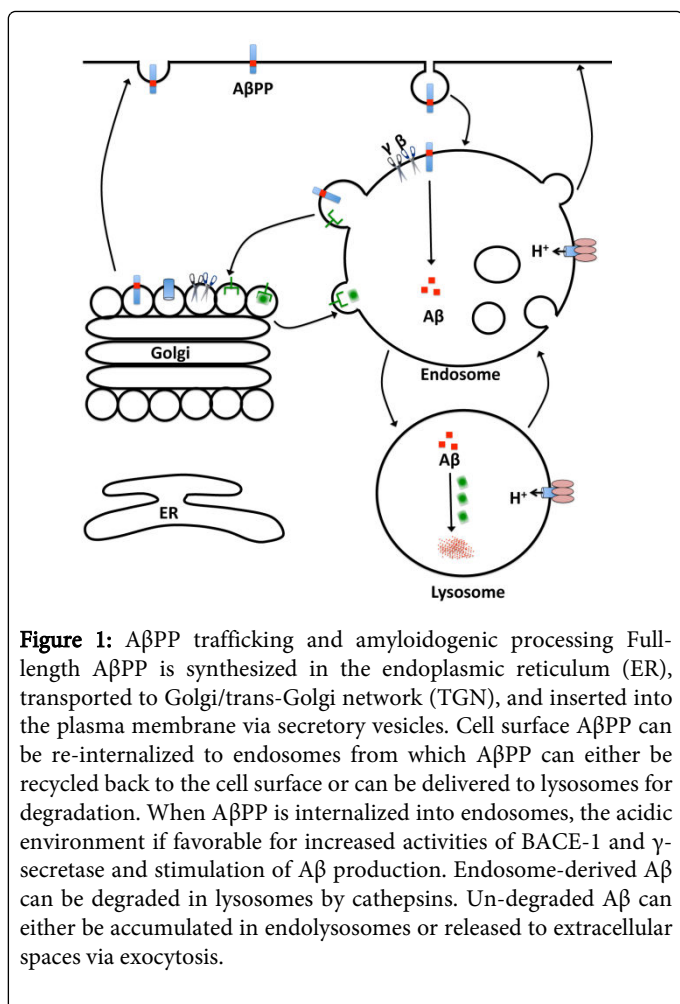
Alzheimer's disease (AD), the most common neurodegenerative disorder of old age, is characterized clinically by a progressive decline in cognitive function, and pathologically by loss of synaptic integrity, loss of neurons and the presence of amyloid plaques composed of amyloid beta (A β) protein and neuronal tangles composed of hyperphosphorylated tau [1,2]. Brain deposition of amyloid beta protein (A β), a proteolytic cleavage product of amyloid beta precursor protein (A β PP) by the beta-site A β PP cleavage enzyme 1 (BACE1) and γ -secretase, continues to be considered an important pathogenic factor of AD [1, 2]. As such, gene mutations in A β PP and presenilin-1 can lead to relatively rare familial AD with early-onset [1]. However, the majority of AD cases is sporadic in nature and is late in onset. Currently, pathogenic mechanisms responsible for sporadic AD remain unclear, but are believed to result from complex interactions between nutritional, environmental, epigenetic and genetic factors [3]. Among these factors, elevated plasma LDL cholesterol represents a robust risk factor for AD pathogenesis. Here, we will discuss the amyloidogenic processing of A β PP, briefly describe cholesterol homeostasis in the periphery and in the brain, discuss the linkage between elevated levels of plasma LDL cholesterol and AD pathogenesis, and explore the underlying mechanisms with a focus on amyloidogenic processing of A β PP.

Amyloidogenic processing of A β PP

Full-length A β PP, a ubiquitously expressed type-I transmembrane protein with largely uncharacterized cellular functions is synthesized in the endoplasmic reticulum and is transported to the Golgi/trans-Golgi network apparatus, where it undergoes posttranslational

modifications and maturation. Once inserted into plasma membranes via secretory vesicles, A β PP can traffic into endosomes via clathrin-dependent endocytosis whereupon it can either be recycled back to the cell surface or delivered to lysosomes for possible degradation [4,5].

Trafficking of A β PP into endolysosomes appears to play a critical role in determining the extent to which A β PP metabolism is non-amyloidogenic or is amyloidogenic [4,6,7]. For the non-amyloidogenic pathway, A β PP in plasma membranes is cleaved by α -secretase to produce sAPP α that is both neurotrophic and neuroprotective [8]. For the amyloidogenic pathway, once A β PP is internalized into the acidic environment of endolysosomes, amyloidogenic metabolism of A β PP is catalyzed by BACE-1 and γ -secretase [9-12]. Amyloidogenesis of endosome-derived A β is further influenced by the ability of A β degradation to be catalyzed by lysosome-resident cathepsins [13]. Remaining levels of A β can either accumulate in endolysosomes as intraneuronal A β or it can be undergo exocytotic release into extracellular spaces where diffuse A β plaque can form (Figure 1). Thus, amyloidogenesis can be enhanced by such factors as those that promote A β PP internalization [14], those that enhance protein levels and/or activities of BACE-1 and/or γ -secretase [15,16], those that prevent A β PP recycling back to the cell surface [17], and those that impair A β degradation in lysosomes [18].



Cholesterol homeostasis in the periphery and in the brain

Cholesterol, an essential component of cellular membranes, helps maintain such physiologically important neuronal functions as neurotransmitter release, neurite outgrowth, and synaptic plasticity [19-21]. Whether synthesized in brain or elsewhere throughout the body, cholesterol is the same chemically. However, there are differences in terms of its association with lipoprotein particles. Lipoproteins vary in size, lipid composition, and complex apolipoproteins that mediate their transport and uptake are different. In plasma, LDL is the main lipoprotein particle that mediates the transport of cholesterol and lipids into peripheral tissues, whereas HDL is the main lipoprotein particle that mediates the reverse cholesterol transport from peripheral tissues. LDL is a 20-25 nm sized particle that has the highest cholesterol content and apoB-100 is the exclusive apolipoprotein that mediates its transport and uptake. HDL, a protein-rich disc-shaped particles, is about 8-10 nm in size, has lower cholesterol content, and primary apolipoproteins that mediate its transport and uptake are apoA-I, apoC-I, apoC-II and apoE.

Brain is the most cholesterol rich organ in the body and contains about 20% of the body's total cholesterol. About 70% of brain cholesterol lies in the myelin sheaths of oligodendroglia and membranes of astrocytes; cholesterol in neurons make up the rest. In

contrast to plasma cholesterol, essentially all cholesterol in the brain is unesterified free cholesterol [22]. Such unesterified free cholesterol is of particular importance to neurons because neurons are extraordinarily polarized cells with extensive processes that require constant membrane trafficking and free cholesterol recycling to maintain physiologically important neuronal functions, including neurotransmitter release, neurite outgrowth, and synaptic plasticity [23,24].

Because the blood-brain barrier (BBB) restricts plasma lipoproteins, especially the larger LDL particles, from entering brain parenchyma, brain cholesterol is almost completely dependent on in situ synthesis of apoE-cholesterol by astrocytes [25]. As such, apoB, the major LDL cholesterol carrier protein in circulating blood, is not present in normal brain [26]. Although the structure and composition of apoE-cholesterol in brain parenchyma is not known, it is estimated that apoE-cholesterol synthesized in situ in brain is a discoidal shaped HDL-like particle composed of phospholipids and unesterified cholesterol, and such an estimation is based on the studies of astrocyte-derived lipoproteins and lipoproteins isolated from the CSF [27,28].

HDL-like apoE-cholesterol synthesized in brain supplies the neuronal need of cholesterol via receptor-mediated endocytosis, via a mechanism involving the Niemann-Pick type C (NPC) proteins type-1 (NPC1) and -2 (NPC2) proteins. Thus, similar to that of plasma HDL, brain in situ synthesized HDL-like apoE-cholesterol may mediate recycling and reverse cholesterol transport [27], and such a function is especially important for fundamental physiological functions of neurons. Indeed, apoE is important for the regulation of synapse formation, plasticity and repair [39, 40] and apoE cholesterol, the nature source of neuronal cholesterol, is neuroprotective [41,42]. Similarly, HDL is neuroprotective [43-45].

Altered cholesterol homeostasis and sporadic AD

Altered cholesterol homeostasis in general and elevation of plasma LDL cholesterol more specifically represents a robust risk factor for AD pathogenesis [46]. This increased risk for AD onset and severity comes from various studies including findings that the presence of the APOE4 allele is still the single strongest genetic risk factor for sporadic AD [47-50], and apoE, the product of the APOE4 gene, is a carrier protein for cholesterol and lipid transport. In plasma, apoE, which is mainly synthesized by the liver and by macrophages and is associated with VLDL, chylomicron remnants, and a subset of HDL particles, plays an important role in reverse cholesterol transport [51,52]. In brain, apoE is synthesized in astrocytes and functions as a cholesterol transport protein between astrocytes and neurons. Although several hypotheses (A β -dependent and A β -independent) have been proposed [34,53-55], the exact underlying mechanisms whereby apoE4 contributes to the pathogenesis of AD remains unclear.

ApoE4 is clearly associated with elevated levels of LDL cholesterol and decreased levels of HDL cholesterol [56,57]. In addition, elevated levels of plasma LDL cholesterol, independent of APOE genotypes, are also linked robustly to the pathogenesis of AD [58-63]. Epidemiologically, elevated levels of LDL cholesterol are associated with increased levels of A β deposition in brain [61] and an increased risk of developing AD [58,63]. Such findings in humans were supported by data from animal studies conducted with A β PP transgenic mice [64,65], guinea pigs [66], rabbits [59,67], and rats [68] - elevated levels of LDL cholesterol leads to memory deficits and

promotes the development of AD-like pathology including brain deposition of A β and/or tau pathology. Similarly, and again independent of the APOE genotype, low levels of HDL cholesterol are also associated with increased risk of developing AD. On the contrary, high levels of HDL cholesterol appear to protect against the occurrence of AD [61,63,69,70]. Thus, altered circulating cholesterol homeostasis, independent of APOE genotype status, is associated with the pathogenesis of sporadic AD. Next, we will explore the underlying mechanisms whereby altered cholesterol homeostasis in the periphery contribute to the pathogenesis of AD in brain with a focus on how elevated levels of plasma LDL cholesterol promote amyloidogenesis.

Plasma LDL cholesterol induces neuroinflammation

Mounting evidence supports the pathogenic role of an inflammatory cascade mediated by activated microglia and reactive astrocytes in the pathogenesis of AD [71-73]. In AD brain, neuroinflammation is deemed to be a secondary response and the likely sources of inflammation are brain deposition of A β , the formation of neurofibrillary tangles, or other neuronal insults and injury [73,74]. Elevated levels of LDL cholesterol have been shown to promote neuroinflammatory responses [65,75]. Although cholesterol-induced brain deposition of A β might be a possible source of neuroinflammation, it is also likely that cholesterol-induced neuroinflammation is a consequence of cholesterol-induced cerebral vascular damage [76]. Such a notion is supported by findings that elevated levels of plasma cholesterol disrupt the BBB [75], an early pathological feature of sporadic AD [77-83], thus allowing systemic macrophages to invade into the brain parenchyma thereby initiating a cascade of neuroinflammatory responses including microglia activation and the formation of reactive astrocytes [65,75]. Such cholesterol-induced neuroinflammation not only could lead to synaptic dysfunction [65] thus contributing to the development of mild cognitive impairment [84], but also could lead to enhanced amyloidogenesis by up-regulation of BACE-1 [65,85] thus contributing to the development of AD, especially when BACE-1 expression is subject to regulation by cytokines [15,16].

Plasma LDL cholesterol induces neuronal endolysosome dysfunction

Under conditions when and where the BBB is disrupted, as occurs early in sporadic AD [77-83], LDL cholesterol can enter brain parenchyma and contribute directly to enhanced amyloidogenesis. Indeed, apoB-100, the exclusive apolipoprotein of LDL-cholesterol thus a marker of peripheral-derived cholesterol [59], is present in AD brain and co-localizes A β [26,59,86-88]. Indeed, we have shown that rabbits fed a diet enriched in cholesterol exhibit elevated levels of LDL cholesterol, disruptions in the integrity of the BBB, and increased brain levels of apoB-100 [59,75]. Once it enters brain, can be internalized by neurons via receptor-mediated endocytosis with the assistance of highly expressed receptors. Because some of these receptors for cholesterol uptake, including LRP1 and LRP10, have been shown to interact with A β PP and affect A β PP trafficking [4,89,90], LDL cholesterol internalization could promote A β PP internalization into neuronal endolysosomes and enhance amyloidogenic processing of A β PP. In support, we have shown that LDL cholesterol treatment promotes A β PP internalization and increases A β accumulation in endolysosomes of primary cultured neurons [60].

Because apoB and apoE have different affinities for receptors for cholesterol uptake, neuronal uptake of apoB-containing LDL cholesterol may result in drastic difference in intracellular cholesterol transport and distribution than that of apoE cholesterol. Additionally, while apoB leads to cholesterol being targeted by the lysosome degradation pathway [91,92], apoE mediates cholesterol recycling [93-95]. Thus, neuronal uptake of apoB-containing LDL cholesterol may lead to cholesterol accumulation in endolysosomes thereby disturbing endolysosome structure and function, a very early pathological feature of AD [9 96-99]. This concept is supported experimentally by findings by others and us that LDL cholesterol treatment increases cholesterol accumulation in neuronal endolysosomes and leads to endolysosome enlargement, elevation of endolysosome pH, and reduced endolysosome enzyme activities [60].

Because endolysosomes are the sites at which internalized A β PP cleavage to A β is catalyzed by BACE-1 and γ -secretase [9-12], and because lysosomes are the sites A β can be degraded by cathepsins [13], disturbed endolysosome structure and function could lead to intraneuronal A β accumulation, another early pathological feature of AD [100-102]. Indeed, we found that LDL cholesterol treatment of neurons increased endolysosome accumulation of BACE-1, enhanced BACE-1 activity, decreased cathepsin activity, and increased endolysosome accumulation of A β [60].

Thus, elevated levels of circulating apoB-containing LDL cholesterol could lead to disturbed intracellular trafficking and distribution of cholesterol that resembles lysosomal lipid storage disorders such as Niemann-Pick type C disease [103-105]. In Niemann-Pick type C disease, the accumulation of cholesterol in lysosomes results in reduced recycling of cholesterol back to ER, Golgi, and plasma membranes thus leading to cholesterol deficiency at sites where it is needed for membrane repair, neurite outgrowth, and synaptic plasticity [39,40]. Moreover, endolysosome accumulation of cholesterol leads to endolysosome dysfunction, which contributes directly to the development of pathological hallmarks of AD including A β deposition, formation of neurofibrillary tangles, and synaptic and neuronal loss [104,106]. It is interesting to note that the association of cholesterol with different apoE isoforms can also result in drastic differences in endocytic trafficking of cholesterol, where apoE4 results in impaired cholesterol recycling [94,107,108]. Such impaired recycling of apoE4-cholesterol in neurons can lead to the accumulation of cholesterol in endolysosomes [94,108], alter endocytic trafficking of A β PP, enhance amyloidogenic processing of A β PP [109], and impair synaptic plasticity [95]. Thus, similar to the effects of apoB-containing LDL cholesterol, the presence of apoE4 can result in decreased cholesterol transport to the plasma membrane and cholesterol deficiency in plasma membranes as well as increased cholesterol accumulation in endolysosomes and subsequent endolysosome dysfunction [97,110], a set of conditions similar to those in Niemann-Pick type C disease albeit less severely.

Under such conditions, the use of statins, a class of hydroxymethylglutaryl-CoA (HMG-CoA) reductase inhibitors that block cholesterol biosynthesis in the ER, would decrease cholesterol transport to plasma membranes thus worsening cholesterol deficits at the plasma membrane, synaptic disruption, and the ability to repair membranes once injured [19,21,111]. In addition, chronic use of statins results in over-expression of LDLRs and enhanced cholesterol uptake [112], and such an effect could increase the cholesterol burden in endolysosomes and worsen endolysosome dysfunction. As such, lipophilic statins, especially those that can cross the BBB and

effectively penetrate cell membranes, can reduce cholesterol synthesis below a critical level that induces neuronal death [113], whereas treatment with hydrophilic statins that do not cross the BBB easily may be appropriate for AD to reduce plasma LDL cholesterol levels without further disturbing neuronal cholesterol homeostasis [114]. Such a notion is supported by findings that statins have no beneficial effects on Niemann-Pick type C disease [103,115]. Such a notion is also supported by recent data and meta-analysis from randomized clinical trials showing that statins have little or no beneficial effects against AD [116-120] and in some cases statins result in adverse effects on memory and cognitions [121-124].

Conclusions

Elevated levels of circulating cholesterol, independent of APOE genotype, appear to contribute to the development of AD. On one hand, elevated levels of plasma LDL cholesterol could promote cerebral vascular damage thus initiating neuroinflammatory responses that contribute to the pathogenesis of AD. On the other hand, LDL cholesterol could disturb neuronal endolysosome function and contribute directly to the pathogenesis AD. Here, we propose a hypothesis that elevated levels of LDL cholesterol lead to lysosome cholesterol storage similar to Niemann-Pick type C disease thus contributing the pathogenesis of AD. Specifically, we propose that plasma LDL cholesterol once it enters brain parenchyma can be internalized by neurons via receptor-mediated endocytosis and can promote A β PP internalization because LDLRs and A β PP physically associate with each other. Unlike apoE-cholesterol, increased apoB-containing LDL-cholesterol could lead to cholesterol accumulation in endolysosomes thus elevating endolysosome pH and impairing endolysosome function. Elevation of endolysosome pH on one hand could lead to increased BACE-1 protein levels and enhanced BACE-1 activity that leads to amyloidogenic processing of A β PP, and on the other hand could reduce cathepsin activity thus impairing A β degradation in lysosomes, thus leading to intraneuronal A β accumulation (Figure 2). Although findings from our animal studies support such a hypothesis, further studies conducted in humans are warranted. It also should be noted that cholesterol is not the only component of LDL particles and associated apolipoproteins, sphingolipids and phospholipids could also affect amyloidogenic processing of A β PP and contribute to the development of sporadic AD [125-128].

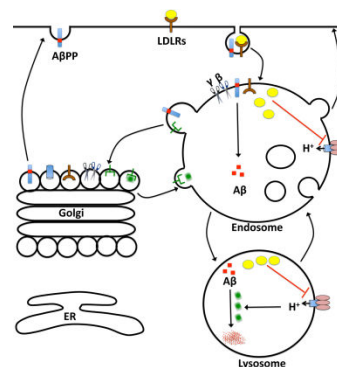


Figure 2: Proposed model of LDL cholesterol-induced amyloidogenic processing of A β PP. Elevated plasma LDL cholesterol, once it enters brain, can be internalized by neurons via receptor-mediated endocytosis. The LDL cholesterol internalization process promotes A β PP internalization. Increased apoB-containing LDL-cholesterol can lead to cholesterol accumulation in endolysosomes thus elevating endolysosome pH and impairing endolysosome function. Elevation of endolysosome pH can lead to increased BACE-1 protein levels and enhanced BACE-1 activity that leads to amyloidogenic processing of APP, and can reduce cathepsin activity thus impairing A β degradation in lysosomes.

Acknowledgements

The authors acknowledge grant support for our studies referred to in this manuscript including P30GM103329, R21AG103329 and R01MH100972.

References

1. Goate A, Hardy J (2012) Twenty years of Alzheimer's disease-causing mutations. See comment in PubMed Commons below *J Neurochem* 120 Suppl 1: 3-8.
2. Holtzman DM, Morris JC, Goate AM (2011) Alzheimer's disease: the challenge of the second century. See comment in PubMed Commons below *Sci Transl Med* 3: 77sr1.
3. Reitz C, Brayne C, Mayeux R (2011) Epidemiology of Alzheimer disease. See comment in PubMed Commons below *Nat Rev Neurol* 7: 137-152.
4. Jiang S, Li Y, Zhang X, Bu G, Xu H, et al. (2014) Trafficking regulation of proteins in Alzheimer's disease. See comment in PubMed Commons below *Mol Neurodegener* 9: 6.
5. Haass C, Kaether C, Thinakaran G, Sisodia S (2012) Trafficking and proteolytic processing of APP. See comment in PubMed Commons below *Cold Spring Harb Perspect Med* 2: a006270.
6. Rajendran L, Annaert W (2012) Membrane trafficking pathways in Alzheimer's disease. See comment in PubMed Commons below *Traffic* 13: 759-770.
7. Morel E, Chamoun Z, Lasiacka ZM, Chan RB, Williamson RL, et al. (2013) Phosphatidylinositol-3-phosphate regulates sorting and processing of amyloid precursor protein through the endosomal system. See comment in PubMed Commons below *Nat Commun* 4: 2250.
8. Thornton E, Vink R, Blumberg PC, and Van Den Heuvel C (2006). Soluble amyloid precursor protein alpha reduces neuronal injury and improves functional outcome following diffuse traumatic brain injury in rats. *Brain Res* 1094: 38-46.

9. Nixon RA (2005) Endosome function and dysfunction in Alzheimer's disease and other neurodegenerative diseases. See comment in PubMed Commons below *Neurobiol Aging* 26: 373-382.
10. Rajendran L, Schneider A, Schlechtingen G, Weidlich S, Ries J, et al. (2008) Efficient inhibition of the Alzheimer's disease beta-secretase by membrane targeting. See comment in PubMed Commons below *Science* 320: 520-523.
11. Sannerud R, Declerck I, Peric A, Raemaekers T, Menendez G, et al. (2011) ADP ribosylation factor 6 (ARF6) controls amyloid precursor protein (APP) processing by mediating the endosomal sorting of BACE1. See comment in PubMed Commons below *Proc Natl Acad Sci U S A* 108: E559-568.
12. Shimizu H, Tosaki A, Kaneko K, Hisano T, Sakurai T, et al. (2008) Crystal structure of an active form of BACE1, an enzyme responsible for amyloid beta protein production. See comment in PubMed Commons below *Mol Cell Biol* 28: 3663-3671.
13. Miners JS, Barua N, Kehoe PG, Gill S, Love S (2011) A β -degrading enzymes: potential for treatment of Alzheimer disease. See comment in PubMed Commons below *J Neuropathol Exp Neurol* 70: 944-959.
14. Grbovic OM, Mathews PM, Jiang Y, Schmidt SD, Dinakar R, et al. (2003). Rab5-stimulated up-regulation of the endocytic pathway increases intracellular beta-cleaved amyloid precursor protein carboxyl-terminal fragment levels and Abeta production. *J Biol Chem* 278: 31261-31268.
15. Sastre M, Dewachter I, Rossner S, Bogdanovic N, Rosen E, et al. (2006) Nonsteroidal anti-inflammatory drugs repress beta-secretase gene promoter activity by the activation of PPARgamma. See comment in PubMed Commons below *Proc Natl Acad Sci U S A* 103: 443-448.
16. Sastre M, Klockgether T, Heneka MT (2006) Contribution of inflammatory processes to Alzheimer's disease: molecular mechanisms. See comment in PubMed Commons below *Int J Dev Neurosci* 24: 167-176.
17. Ma QL, Galasko DR, Ringman JM, Vinters HV, Edland SD, et al. (2009) Reduction of SorLA/LR11, a sorting protein limiting beta-amyloid production, in Alzheimer disease cerebrospinal fluid. See comment in PubMed Commons below *Arch Neurol* 66: 448-457.
18. Torres M, Jimenez S, Sanchez-Varo R, Navarro V, Trujillo-Estrada L, et al. (2012) Defective lysosomal proteolysis and axonal transport are early pathogenic events that worsen with age leading to increased APP metabolism and synaptic Abeta in transgenic APP/PS1 hippocampus. *Mol Neurodegener* 7: 59.
19. Linetti A, Fratangeli A, Taverna E, Valnegri P, Francolini M, et al. (2010) Cholesterol reduction impairs exocytosis of synaptic vesicles. See comment in PubMed Commons below *J Cell Sci* 123: 595-605.
20. Koudinov AR, Koudinova NV (2001) Essential role for cholesterol in synaptic plasticity and neuronal degeneration. See comment in PubMed Commons below *FASEB J* 15: 1858-1860.
21. Fünfschilling U, Jockusch WJ, Sivakumar N, Möbius W, Corthals K, et al. (2012) Critical time window of neuronal cholesterol synthesis during neurite outgrowth. See comment in PubMed Commons below *J Neurosci* 32: 7632-7645.
22. Björkhem I, Meaney S (2004) Brain cholesterol: long secret life behind a barrier. See comment in PubMed Commons below *Arterioscler Thromb Vasc Biol* 24: 806-815.
23. Orth M, Bellosta S (2012) Cholesterol: its regulation and role in central nervous system disorders. See comment in PubMed Commons below *Cholesterol* 2012: 292598.
24. Vance JE, Hayashi H, Karten B (2005) Cholesterol homeostasis in neurons and glial cells. See comment in PubMed Commons below *Semin Cell Dev Biol* 16: 193-212.
25. Nieweg K, Schaller H, Pfrieger FW (2009) Marked differences in cholesterol synthesis between neurons and glial cells from postnatal rats. See comment in PubMed Commons below *J Neurochem* 109: 125-134.
26. Pitas RE, Boyles JK, Lee SH, Hui D, Weisgraber KH (1987) Lipoproteins and their receptors in the central nervous system. Characterization of the lipoproteins in cerebrospinal fluid and identification of apolipoprotein B,E(LDL) receptors in the brain. See comment in PubMed Commons below *J Biol Chem* 262: 14352-14360.
27. de Chaves EP, Narayanaswami V (2008) Apolipoprotein E and cholesterol in aging and disease in the brain. See comment in PubMed Commons below *Future Lipidol* 3: 505-530.
28. Vance JE (2012) Dysregulation of cholesterol balance in the brain: contribution to neurodegenerative diseases. See comment in PubMed Commons below *Dis Model Mech* 5: 746-755.
29. Maxfield FR, Tabas I (2005) Role of cholesterol and lipid organization in disease. See comment in PubMed Commons below *Nature* 438: 612-621.
30. Vance JE, Karten B, Hayashi H (2006) Lipid dynamics in neurons. See comment in PubMed Commons below *Biochem Soc Trans* 34: 399-403.
31. Sleat DE, Wiseman JA, El-Banna M, Price SM, Verot L, et al. (2004) Genetic evidence for nonredundant functional cooperativity between NPC1 and NPC2 in lipid transport. See comment in PubMed Commons below *Proc Natl Acad Sci U S A* 101: 5886-5891.
32. Beffert U, Danik M, Krzywkowski P, Ramassamy C, Berrada F, et al. (1998) The neurobiology of apolipoproteins and their receptors in the CNS and Alzheimer's disease. See comment in PubMed Commons below *Brain Res Brain Res Rev* 27: 119-142.
33. Dietschy JM (2009) Central nervous system: cholesterol turnover, brain development and neurodegeneration. See comment in PubMed Commons below *Biol Chem* 390: 287-293.
34. Liu CC, Kanekiyo T, Xu H, Bu G (2013) Apolipoprotein E and Alzheimer disease: risk, mechanisms and therapy. See comment in PubMed Commons below *Nat Rev Neurol* 9: 106-118.
35. Holtzman DM, Herz J, Bu G (2012) Apolipoprotein E and apolipoprotein E receptors: normal biology and roles in Alzheimer disease. See comment in PubMed Commons below *Cold Spring Harb Perspect Med* 2: a006312.
36. Coraci IS, Husemann J, Berman JW, Hulette C, Dufour JH, et al. (2002) CD36, a class B scavenger receptor, is expressed on microglia in Alzheimer's disease brains and can mediate production of reactive oxygen species in response to beta-amyloid fibrils. See comment in PubMed Commons below *Am J Pathol* 160: 101-112.
37. Husemann J, Loike JD, Anankov R, Febbraio M, Silverstein SC (2002) Scavenger receptors in neurobiology and neuropathology: their role on microglia and other cells of the nervous system. See comment in PubMed Commons below *Glia* 40: 195-205.
38. Lucarelli M, Borrelli V, Fiori A, Cucina A, Granata F, et al. (2002) The expression of native and oxidized LDL receptors in brain microvessels is specifically enhanced by astrocytes-derived soluble factor(s). See comment in PubMed Commons below *FEBS Lett* 522: 19-23.
39. Rebeck GW, Kindy M, LaDu MJ (2002) Apolipoprotein E and Alzheimer's disease: the protective effects of ApoE2 and E3. See comment in PubMed Commons below *J Alzheimers Dis* 4: 145-154.
40. Poirier J (2008) Apolipoprotein E represents a potent gene-based therapeutic target for the treatment of sporadic Alzheimer's disease. See comment in PubMed Commons below *Alzheimers Dement* 4: S91-97.
41. Hayashi H, Eguchi Y, Fukuchi-Nakaishi Y, Takeya M, Nakagata N, et al. (2012). A potential neuroprotective role of apolipoprotein E-containing lipoproteins through low density lipoprotein receptor-related protein 1 in normal tension glaucoma. *J Biol Chem* 287: 25395-25406.
42. Hayashi H, Campenot RB, Vance DE, Vance JE (2007). Apolipoprotein E-containing lipoproteins protect neurons from apoptosis via a signaling pathway involving low-density lipoprotein receptor-related protein-1. *J Neurosci* 27: 1933-1941.
43. Kontush A, Chapman MJ (2008) HDL: close to our memories? See comment in PubMed Commons below *Arterioscler Thromb Vasc Biol* 28: 1418-1420.
44. Paternò R, Ruocco A, Postiglione A, Hubsch A, Andresen I, et al. (2004) Reconstituted high-density lipoprotein exhibits neuroprotection in two rat models of stroke. See comment in PubMed Commons below *Cerebrovasc Dis* 17: 204-211.
45. Lapergue B, Moreno JA, Dang BQ, Coutard M, Delbosc S, et al. (2010) Protective effect of high-density lipoprotein-based therapy in a model of

- embolic stroke. See comment in PubMed Commons below *Stroke* 41: 1536-1542.
46. Fiorenza MT, Dardis A2, Canterini S1, Erickson RP3 (2013) Cholesterol metabolism-associated molecules in late onset Alzheimer's disease. See comment in PubMed Commons below *J Biol Regul Homeost Agents* 27: 23-35.
47. Reitz C, Rogaeva E, Foroud T, Farrer LA (2011) Genetics and genomics of late-onset Alzheimer's disease and its endophenotypes. See comment in PubMed Commons below *Int J Alzheimers Dis* 2011: 284728.
48. Farrer LA, Cupples LA, Haines JL, Hyman B, Kukull WA, et al. (1997) Effects of age, sex, and ethnicity on the association between apolipoprotein E genotype and Alzheimer disease. A meta-analysis. APOE and Alzheimer Disease Meta Analysis Consortium. See comment in PubMed Commons below *JAMA* 278: 1349-1356.
49. Harold D, Abraham R, Hollingworth P, Sims R, Gerrish A, et al. (2009) Genome-wide association study identifies variants at CLU and PICALM associated with Alzheimer's disease. See comment in PubMed Commons below *Nat Genet* 41: 1088-1093.
50. Lambert JC, Heath S, Even G, Campion D, Sleegers K, et al. (2009) Genome-wide association study identifies variants at CLU and CR1 associated with Alzheimer's disease. See comment in PubMed Commons below *Nat Genet* 41: 1094-1099.
51. Mahley RW, Huang Y, Weisgraber KH (2006) Putting cholesterol in its place: apoE and reverse cholesterol transport. See comment in PubMed Commons below *J Clin Invest* 116: 1226-1229.
52. Matsuura F, Wang N, Chen W, Jiang XC, Tall AR (2006) HDL from CETP-deficient subjects shows enhanced ability to promote cholesterol efflux from macrophages in an apoE- and ABCG1-dependent pathway. See comment in PubMed Commons below *J Clin Invest* 116: 1435-1442.
53. Vergheze PB, Castellano JM, Garai K, Wang Y, Jiang H, et al. (2013) ApoE influences amyloid- β^2 ($A\beta^2$) clearance despite minimal apoE/ $A\beta^2$ association in physiological conditions. See comment in PubMed Commons below *Proc Natl Acad Sci U S A* 110: E1807-1816.
54. Kanekiyo T, Xu H2, Bu G3 (2014) ApoE and $A\beta^2$ in Alzheimer's disease: accidental encounters or partners? See comment in PubMed Commons below *Neuron* 81: 740-754.
55. Tai LM, Mehra S, Shete V, Estus S, Rebeck GW, et al. (2014) Soluble apoE/ $A\beta^2$ complex: mechanism and therapeutic target for APOE4-induced AD risk. See comment in PubMed Commons below *Mol Neurodegener* 9: 2.
56. Corder EH, Saunders AM, Strittmatter WJ, Schmechel DE, Gaskell PC, et al. (1993) Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. See comment in PubMed Commons below *Science* 261: 921-923.
57. Marzolo MP, Bu G (2009) Lipoprotein receptors and cholesterol in APP trafficking and proteolytic processing, implications for Alzheimer's disease. See comment in PubMed Commons below *Semin Cell Dev Biol* 20: 191-200.
58. Solomon A, Kivipelto M, Wolozin B, Zhou J, Whitmer RA (2009) Midlife serum cholesterol and increased risk of Alzheimer's and vascular dementia three decades later. See comment in PubMed Commons below *Dement Geriatr Cogn Disord* 28: 75-80.
59. Chen X, Wagener JF, Morgan DH, Hui L, Ghribi O, et al. (2010) Endolysosome mechanisms associated with Alzheimer's disease-like pathology in rabbits ingesting cholesterol-enriched diet. See comment in PubMed Commons below *J Alzheimers Dis* 22: 1289-1303.
60. Hui L, Chen X, Geiger JD (2012) Endolysosome involvement in LDL cholesterol-induced Alzheimer's disease-like pathology in primary cultured neurons. See comment in PubMed Commons below *Life Sci* 91: 1159-1168.
61. Reed B, Villeneuve S2, Mack W3, DeCarli C1, Chui HC3, et al. (2014) Associations between serum cholesterol levels and cerebral amyloidosis. See comment in PubMed Commons below *JAMA Neurol* 71: 195-200.
62. Matsuzaki T, Sasaki K, Hata J, Hirakawa Y, Fujimi K, et al. (2011) Association of Alzheimer disease pathology with abnormal lipid metabolism: the Hisayama Study. See comment in PubMed Commons below *Neurology* 77: 1068-1075.
63. Kuo YM, Emmerling MR, Bisgaier CL, Essenburg AD, Lampert HC, et al. (1998) Elevated low-density lipoprotein in Alzheimer's disease correlates with brain abeta 1-42 levels. See comment in PubMed Commons below *Biochem Biophys Res Commun* 252: 711-715.
64. Barbero-Camps E, Fernandez A, Martinez L, Fernandez-Checa JC, Colell A (2013). APP/PS1 mice overexpressing SREBP-2 exhibit combined Abeta accumulation and tau pathology underlying Alzheimer's disease. *Hum Mol Genet* 22: 3460-3476.
65. Thirumangalakudi L, Prakasam A, Zhang R, Bimonte-Nelson H, Sambamurti K, et al. (2008) High cholesterol-induced neuroinflammation and amyloid precursor protein processing correlate with loss of working memory in mice. *J Neurochem* 106: 475-485.
66. Sharman MJ, Moussavi Nik SH, Chen MM, Ong D, Wijaya L, et al. (2013) The Guinea Pig as a Model for Sporadic Alzheimer's Disease (AD): The Impact of Cholesterol Intake on Expression of AD-Related Genes. See comment in PubMed Commons below *PLoS One* 8: e66235.
67. Sparks DL, Scheff SW, Hunsaker JC 3rd, Liu H, Landers T, et al. (1994) Induction of Alzheimer-like beta-amyloid immunoreactivity in the brains of rabbits with dietary cholesterol. See comment in PubMed Commons below *Exp Neurol* 126: 88-94.
68. Ullrich C, Pirchl M, Humpel C (2010) Hypercholesterolemia in rats impairs the cholinergic system and leads to memory deficits. See comment in PubMed Commons below *Mol Cell Neurosci* 45: 408-417.
69. Reitz C, Tang MX, Schupf N, Manly JJ, Mayeux R et al. (2010). Association of higher levels of high-density lipoprotein cholesterol in elderly individuals and lower risk of late-onset Alzheimer disease. *Arch Neurol* 67: 1491-1497.
70. Yasuno F, Asada T (2013) Effect of plasma lipids and APOE genotype on cognitive decline. See comment in PubMed Commons below *Dialogues Clin Neurosci* 15: 120-126.
71. Heneka MT, O'Banion MK (2007) Inflammatory processes in Alzheimer's disease. See comment in PubMed Commons below *J Neuroimmunol* 184: 69-91.
72. Hensley K (2010) Neuroinflammation in Alzheimer's disease: mechanisms, pathologic consequences, and potential for therapeutic manipulation. See comment in PubMed Commons below *J Alzheimers Dis* 21: 1-14.
73. Morales I, Guzmán-Martínez L1, Cerda-Troncoso C1, Fariás GA2, Maccioni RB3 (2014) Neuroinflammation in the pathogenesis of Alzheimer's disease. A rational framework for the search of novel therapeutic approaches. See comment in PubMed Commons below *Front Cell Neurosci* 8: 112.
74. Hardy J, Selkoe DJ (2002) The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. See comment in PubMed Commons below *Science* 297: 353-356.
75. Chen X, Gawryluk JW, Wagener JF, Ghribi O, Geiger JD (2008) Caffeine blocks disruption of blood brain barrier in a rabbit model of Alzheimer's disease. See comment in PubMed Commons below *J Neuroinflammation* 5: 12.
76. Perry VH, Newman TA, Cunningham C (2003) The impact of systemic infection on the progression of neurodegenerative disease. See comment in PubMed Commons below *Nat Rev Neurosci* 4: 103-112.
77. Zipser BD, Johanson CE, Gonzalez L, Berzin TM, Tavares R, et al. (2007) Microvascular injury and blood-brain barrier leakage in Alzheimer's disease. See comment in PubMed Commons below *Neurobiol Aging* 28: 977-986.
78. Ujiié M, Dickstein DL, Carlow DA, and Jefferies WA (2003). Blood-brain barrier permeability precedes senile plaque formation in an Alzheimer disease model. *Microcirculation* 10: 463-470.
79. Kalaria RN (1999) The blood-brain barrier and cerebrovascular pathology in Alzheimer's disease. See comment in PubMed Commons below *Ann N Y Acad Sci* 893: 113-125.

80. Kalaria RN (1992) The blood-brain barrier and cerebral microcirculation in Alzheimer disease. See comment in PubMed Commons below *Cerebrovasc Brain Metab Rev* 4: 226-260.
81. Munoz DG, Erkinjuntti T, Gaytan-Garcia S, Hachinski V (1997) Serum protein leakage in Alzheimer's disease revisited. See comment in PubMed Commons below *Ann N Y Acad Sci* 826: 173-189.
82. Bell RD, Winkler EA, Singh I, Sagare AP, Deane R, et al. (2012) Apolipoprotein E controls cerebrovascular integrity via cyclophilin A. See comment in PubMed Commons below *Nature* 485: 512-516.
83. Grammas P, Samany PG, Thirumangalakudi L (2006) Thrombin and inflammatory proteins are elevated in Alzheimer's disease microvessels: implications for disease pathogenesis. See comment in PubMed Commons below *J Alzheimers Dis* 9: 51-58.
84. Hascalovici JR, Vaya J, Khatib S, Holcroft CA, Zukor H, et al. (2009) Brain sterol dysregulation in sporadic AD and MCI: relationship to heme oxygenase-1. See comment in PubMed Commons below *J Neurochem* 110: 1241-1253.
85. Ghribi O, Larsen B, Schrag M, Herman MM (2006) High cholesterol content in neurons increases BACE, beta-amyloid, and phosphorylated tau levels in rabbit hippocampus. See comment in PubMed Commons below *Exp Neurol* 200: 460-467.
86. Namba Y, Tsuchiya H, Ikeda K (1992) Apolipoprotein B immunoreactivity in senile plaque and vascular amyloids and neurofibrillary tangles in the brains of patients with Alzheimer's disease. See comment in PubMed Commons below *Neurosci Lett* 134: 264-266.
87. Takechi R, Galloway S, Pallebage-Gamarallage M, Wellington C, Johnsen R et al. (2009). Three-dimensional colocalization analysis of plasma-derived apolipoprotein B with amyloid plaques in APP/PS1 transgenic mice. *Histochem Cell Biol* 131: 661-666.
88. Berezcki E, Bernát G, Csont T, Ferdinandy P, Scheich H, et al. (2008) Overexpression of human apolipoprotein B-100 induces severe neurodegeneration in transgenic mice. See comment in PubMed Commons below *J Proteome Res* 7: 2246-2252.
89. Brodeur J, Thériault C, Lessard-Beaudoin M, Marciel A, Dahan S, et al. (2012) LDLR-related protein 10 (LRP10) regulates amyloid precursor protein (APP) trafficking and processing: evidence for a role in Alzheimer's disease. See comment in PubMed Commons below *Mol Neurodegener* 7: 31.
90. Yoon IS, Chen E, Busse T, Repetto E, Lakshmana MK et al. (2007). Low-density lipoprotein receptor-related protein promotes amyloid precursor protein trafficking to lipid rafts in the endocytic pathway. *FASEB J* 21: 2742-2752.
91. Laatsch A, Panteli M, Sornsakrin M, Hoffzimmer B, Grewal T, et al. (2012) Low density lipoprotein receptor-related protein 1 dependent endosomal trapping and recycling of apolipoprotein E. See comment in PubMed Commons below *PLoS One* 7: e29385.
92. Rensen PC, Jong MC, van Vark LC, van der Boom H, Hendriks WL, et al. (2000) Apolipoprotein E is resistant to intracellular degradation in vitro and in vivo. Evidence for retroendocytosis. See comment in PubMed Commons below *J Biol Chem* 275: 8564-8571.
93. Heeren J, Grewal T, Laatsch A, Rottke D, Rinninger F, et al. (2003) Recycling of apoprotein E is associated with cholesterol efflux and high density lipoprotein internalization. See comment in PubMed Commons below *J Biol Chem* 278: 14370-14378.
94. Heeren J, Grewal T, Laatsch A, Becker N, Rinninger F, et al. (2004) Impaired recycling of apolipoprotein E4 is associated with intracellular cholesterol accumulation. See comment in PubMed Commons below *J Biol Chem* 279: 55483-55492.
95. Chen Y, Durakoglugil MS, Xian X, Herz J (2010) ApoE4 reduces glutamate receptor function and synaptic plasticity by selectively impairing ApoE receptor recycling. See comment in PubMed Commons below *Proc Natl Acad Sci U S A* 107: 12011-12016.
96. Yuyama K, Yanagisawa K (2009) Late endocytic dysfunction as a putative cause of amyloid fibril formation in Alzheimer's disease. See comment in PubMed Commons below *J Neurochem* 109: 1250-1260.
97. Cataldo AM, Peterhoff CM, Troncoso JC, Gomez-Isla T, Hyman BT, et al. (2000). Endocytic pathway abnormalities precede amyloid beta deposition in sporadic Alzheimer's disease and Down syndrome: differential effects of APOE genotype and presenilin mutations. *Am J Pathol* 157: 277-286.
98. Tate BA, Mathews PM (2006) Targeting the role of the endosome in the pathophysiology of Alzheimer's disease: a strategy for treatment. See comment in PubMed Commons below *Sci Aging Knowledge Environ* 2006: re2.
99. Boland B, Kumar A, Lee S, Platt FM, Wegiel J, et al. (2008) Autophagy induction and autophagosome clearance in neurons: relationship to autophagic pathology in Alzheimer's disease. See comment in PubMed Commons below *J Neurosci* 28: 6926-6937.
100. Cataldo AM, Petanceska S, Terio NB, Peterhoff CM, Durham R, et al. (2004) Abeta localization in abnormal endosomes: association with earliest Abeta elevations in AD and Down syndrome. See comment in PubMed Commons below *Neurobiol Aging* 25: 1263-1272.
101. Giménez-Llort L, Blázquez G, Cañete T, Johansson B, Oddo S, et al. (2007) Modeling behavioral and neuronal symptoms of Alzheimer's disease in mice: a role for intraneuronal amyloid. See comment in PubMed Commons below *Neurosci Biobehav Rev* 31: 125-147.
102. Bayer TA, Wirths O (2011) Intraneuronal A β as a trigger for neuron loss: can this be translated into human pathology? See comment in PubMed Commons below *Biochem Soc Trans* 39: 857-861.
103. Madra M, Sturley SL (2010) Niemann-Pick type C pathogenesis and treatment: from statins to sugars. See comment in PubMed Commons below *Clin Lipidol* 5: 387-395.
104. Malnar M, Hecimovic S2, Mattsson N3, Zetterberg H4 (2014) Bidirectional links between Alzheimer's disease and Niemann-Pick type C disease. See comment in PubMed Commons below *Neurobiol Dis* .
105. Pacheco CD, Lieberman AP (2008) The pathogenesis of Niemann-Pick type C disease: a role for autophagy? See comment in PubMed Commons below *Expert Rev Mol Med* 10: e26.
106. Pressey SN, Smith DA, Wong AM, Platt FM, Cooper JD (2012) Early glial activation, synaptic changes and axonal pathology in the thalamocortical system of Niemann-Pick type C1 mice. See comment in PubMed Commons below *Neurobiol Dis* 45: 1086-1100.
107. DeKroon RM, Armati PJ (2001) The endosomal trafficking of apolipoprotein E3 and E4 in cultured human brain neurons and astrocytes. See comment in PubMed Commons below *Neurobiol Dis* 8: 78-89.
108. Gong JS, Morita SY, Kobayashi M, Handa T, Fujita SC, et al. (2007). Novel action of apolipoprotein E (ApoE): ApoE isoform specifically inhibits lipid-particle-mediated cholesterol release from neurons. *Mol Neurodegener* 2: 9.
109. Ye S, Huang Y, Mullendorff K, Dong L, Giedt G, et al. (2005). Apolipoprotein (apo) E4 enhances amyloid beta peptide production in cultured neuronal cells: apoE structure as a potential therapeutic target. *Proc Natl Acad Sci U S A* 102: 18700-18705.
110. Nixon RA (2004) Niemann-Pick Type C disease and Alzheimer's disease: the APP-endosome connection fattens up. See comment in PubMed Commons below *Am J Pathol* 164: 757-761.
111. Mailman T, Hariharan M, Karten B (2011) Inhibition of neuronal cholesterol biosynthesis with lovastatin leads to impaired synaptic vesicle release even in the presence of lipoproteins or geranylgeraniol. See comment in PubMed Commons below *J Neurochem* 119: 1002-1015.
112. Goldstein JL, Brown MS (2009) The LDL receptor. See comment in PubMed Commons below *Arterioscler Thromb Vasc Biol* 29: 431-438.
113. Michikawa M, Yanagisawa K (1998) Apolipoprotein E4 induces neuronal cell death under conditions of suppressed de novo cholesterol synthesis. See comment in PubMed Commons below *J Neurosci Res* 54: 58-67.
114. Sparks DL, Connor DJ, Sabbagh MN, Petersen RB, Lopez J, et al. (2006). Circulating cholesterol levels, apolipoprotein E genotype and dementia severity influence the benefit of atorvastatin treatment in Alzheimer's disease: results of the Alzheimer's Disease Cholesterol-Lowering Treatment (ADCLT) trial. *Acta Neurol Scand Suppl* 185: 3-7.

115. Patterson MC, Di Bisceglie AM, Higgins JJ, Abel RB, Schiffmann R, et al. (1993) The effect of cholesterol-lowering agents on hepatic and plasma cholesterol in Niemann-Pick disease type C. See comment in PubMed Commons below *Neurology* 43: 61-64.
116. McGuinness, Craig D, Bullock R, Malouf R, and Passmore P (2014). Statins for the treatment of dementia. *Cochrane Database Syst Rev* 7: CD007514.
117. Silva T, Teixeira J, Remião F, Borges F (2013) Alzheimer's disease, cholesterol, and statins: the junctions of important metabolic pathways. See comment in PubMed Commons below *Angew Chem Int Ed Engl* 52: 1110-1121.
118. Shinohara M, Sato N1, Shimamura M2, Kurinami H2, Hamasaki T3, et al. (2014) Possible modification of Alzheimer's disease by statins in midlife: interactions with genetic and non-genetic risk factors. See comment in PubMed Commons below *Front Aging Neurosci* 6: 71.
119. Wong WB, Lin VW, Boudreau D, Devine EB (2013) Statins in the prevention of dementia and Alzheimer's disease: a meta-analysis of observational studies and an assessment of confounding. See comment in PubMed Commons below *Pharmacoepidemiol Drug Saf* 22: 345-358.
120. Kelley BJ, Glasser S (2014) Cognitive effects of statin medications. See comment in PubMed Commons below *CNS Drugs* 28: 411-419.
121. Orsi A1, Sherman O, Woldelessie Z (2001) Simvastatin-associated memory loss. See comment in PubMed Commons below *Pharmacotherapy* 21: 767-769.
122. Evans MA, Golomb BA (2009) Statin-associated adverse cognitive effects: survey results from 171 patients. See comment in PubMed Commons below *Pharmacotherapy* 29: 800-811.
123. Harrison RW, Ashton CH (1994) Do cholesterol-lowering agents affect brain activity? A comparison of simvastatin, pravastatin, and placebo in healthy volunteers. See comment in PubMed Commons below *Br J Clin Pharmacol* 37: 231-236.
124. Goldstein MR, Mascitelli L, Pezzetta F, Haan MN, Cramer C, et al. (2009) Use of statins and incidence of dementia and cognitive impairment without dementia in a cohort study. See comment in PubMed Commons below *Neurology* 72: 1190-1191.
125. Löffler T, Flunkert S, Havas D, Sántha M, Hutter-Paier B, et al. (2013) Impact of ApoB-100 expression on cognition and brain pathology in wild-type and hAPPsl mice. See comment in PubMed Commons below *Neurobiol Aging* 34: 2379-2388.
126. Mapstone M, Cheema AK2, Fiandaca MS3, Zhong X4, Mhyre TR5, et al. (2014) Plasma phospholipids identify antecedent memory impairment in older adults. See comment in PubMed Commons below *Nat Med* 20: 415-418.
127. Kosicek M, Hecimovic S (2013) Phospholipids and Alzheimer's disease: alterations, mechanisms and potential biomarkers. See comment in PubMed Commons below *Int J Mol Sci* 14: 1310-1322.
128. Mielke MM, Haughey NJ, Bandaru VV, Weinberg DD, Darby E, et al. (2011) Plasma sphingomyelins are associated with cognitive progression in Alzheimer's disease. See comment in PubMed Commons below *J Alzheimers Dis* 27: 259-269.