Joint Destruction and Osteoporosis are Associated with Upregulation of IL-34 and Cathepsin k Expression in Rheumatoid Arthritis. Clinical Trial with Anti TNF α Therapy

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Abstract

Background: Previous studies demonstrated that significant association was found between IL-34 synovial tissue expression and synovitis severity in RA. Furthermore the overexpression of cathepsin K in RA synovia proves that this protease may become a new and highly specific biomarker for RA.

Objective: To find out whether serum levels of IL-34 and Cathepsin-K vary in patients with longstanding RA treated with biologic therapy versus early RA patients treated with conventional DMARDs. Also to estimate any association between their baseline serum levels and disease activity, subsequent joint destruction and osteoporosis.

Methods: This study included forty one RA patients, 21 as a patient group who started treatment with anti TNFα therapy. Group of controls included 20 RA patients who were treated with DMARDs. Full clinical and laboratory assessment as well as radiological by Van Der Heide Sharp score (SHS) and DEXA T scores were done. Serum IL34 and cathepsin k were assessed by ELISA before and one year after treatment.

Results: After one year of therapy, patients group had significant lower serum IL-34 (s.IL-34) level, cathepsin K level CRP, DAS28, DEXA T-score, and SHS (p<0.01) than baseline values(p>0.01), while no significant change in s.IL-34, cathepsin-K in controls. Baseline sIL34 and cathepsin K were positively correlated with DAS28 and SHS and DEXA T scores (p<0.05). There was a significant difference in morning stiffness, DAS28, and serum IL-34 in good responders versus poor responders according to WHO/ILAR response criteria of improvement among patients group. High baseline DAS28 is independent risk factors for radiographic change in RA while high baseline CRP is a risk factor for osteoporosis in RA patients.

Conclusion: Serum IL 34 and cathepsin k were strongly linked to disease activity and duration in RA patients and were highly relevant to both localized osteoporosis and generalized osteoporosis. Also, Anti TNFα therapy effectively decrease both biomarkers regardless drug type, with amelioration of clinical, laboratory and radiological parameters of RA patients.

Keywords: Rheumatoid arthritis; Joint destruction; Osteoporosis; IL34; Cathepsin K; Anti Tnfa drugs

Introduction

Rheumatoid arthritis (RA) is an autoimmune chronic inflammatory disease characterized by inflammatory infiltration of the synovium and synovial hyperplasia, leading to cartilage degradation and bone destruction which may be manifested as erosions, localized juxta-articular bone loss, or generalized bone loss [1]. Because functional outcome in RA is dependent on the extent of joint destruction, early diagnosis and immediate aggressive treatment of RA are required to prevent progressive joint damage, functional disability and reduced quality of life [2].

Osteoclasts (OCs) differentiate from the monocyte/macrophage lineage of hematopoietic myeloid progenitors and its differentiation correlates with the severity of the inflammatory condition. Receptor activator of nuclear factor-κB ligand (RANKL) and macrophage colony-stimulating factor (M-CSF) (also called CSF-1) are considered crucial in osteoclast differentiation and function [3]. Imbalance between osteoblast and osteoclast activities due to inflammatory cytokines pannus interface, periarticular and generalized bone loss in RA pathophysiology [5]. Tumor necrosis factor alpha (TNF-α) is mediating mobilization of osteoclast precursors (OCPs) from bone marrow into the inflamed joint. Also, it stimulates fibroblast-like synovial cells (FLS) to increase cytokines production, which accelerates OC activation in the inflamed synovium of RA [6].

Interleukin–34 (IL-34) is a newly discovered cytokine [7]. It shares a common receptor (c-Fms) with M-CSF, which is expressed on the cell surface of human monocytes [8]. Although both of them share the same receptor, their signal transduction mechanisms and biological activity are not identical [9].

Studies showed that IL-34 can stimulate colony formation of macrophages in human bone marrow cells in a similar efficacy like M-CSF and can substitute entirely for it in RANKL-induced osteoclastogenesis [10]. Chemel et al., demonstrated that significant

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Received August 17, 2015; Accepted September 08, 2015; Published September 19, 2015

Citation: Saber N, Atef M, AbdelAziz D (2015) Joint Destruction and Osteoporosis are Associated with Upregulation of IL-34 and Cathepsin k Expression in Rheumatoid Arthritis. Clinical Trial with Anti TNF α Therapy. J Arthritis 4: 167. doi:10.4172/2167-7921.1000167

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association was found between IL-34 synovial tissue expression and synovitis severity in RA [11]. Eda et al., documented that IL-34 could increase IL-6 and chemokine levels in human whole blood [12]. It also could promote osteoclastogenesis in vitro [13]. Tian et al., found that IL-34 expression could be induced by TNF-α and IL-1β which are pivotal cytokines in RA [14]. Studies have shown that administration of an antibody or inhibitor against the c-Fms selectively and completely blocks osteoclastogenesis and bone erosion induced by TNF-α. Thus, identifying factors involved in TNFα- induced osteoclastogenesis that contribute to erosive arthritis is of great clinical importance [15]. Furthermore, the administration of TNFα blocking agents results in a decrease in the RA inflammatory responses and provides a clinical improvement [16].

Cathepsins (B, L, S, K) are an extensive family of cysteine proteases that are regulated by cytokines and have broad proteolytic activity including activity on types II, IX, and XI collagen and proteoglycans [17]. Cathepsin K is considered to be one of the most important proteolytic enzymes in osteoclastic bone resorption as it can degrade type I collagen at multiple sites in the triple helical domains [18]. Cathepsin K enzyme is abundantly detected in osteoclasts along the bone resorption surfaces, in intracellular lysosomes and transcytotic vesicles [19].

It is expressed by both macrophages and fibroblasts in RA synovial tissue and is present in significantly higher concentrations than in osteoarthritis (OA) [20].

Cathepsins potential role as mediators of bone destruction in arthritis was confirmed in studies in which a cysteine protease inhibitor significantly decreased joint damage in the animal arthritis model [21]. Besides collagens, cathepsin K cleaves a variety of other bone- and cartilage resident proteins such as osteonectin, aggrecan, and IGF-1 [22,23].

In this context, we inquire whether IL-34 and cathepsin K , two different inflammatory mediators have synergistic roles in the pathogenesis and what will be the effect of administration of TNFα inhibitors drugs on localized and generalized osteoporosis.

The objective of the present study is to find out whether serum levels of IL-34 and Cathepsin-K vary in patients with longstanding RA treated with biologic therapy versus early RA patients treated with conventional DMARDS. Also to estimate any association between their baseline serum levels and disease activity, subsequent joint destruction and osteoporosis.

Methods

This study included 41 RA patients presented to outpatient clinic of Physical Medicine, Rheumatology and Rehabilitation Department of Ain Shams University Hospitals.

Inclusion criteria

RA patients fulfilled the American College of Rheumatology (ACR) and ACR/European League Against Rheumatism (EULAR) 2010 criteria for RA [24,25], age > 18 years, disease activity score (DAS28) > 3.2. They were classified into either cases or controls according to their baseline serum levels and disease activity, subsequent joint destruction and osteoporosis. The study was conducted in accordance with the World Medical Association Declaration of Helsinki for human subjects and the study was approved by the ethics committee of the faculty of Medicine and all patients were informed and gave their written consent.

Exclusion criteria

Patients who had Paget disease, multiple myeloma, breast cancer, bone metastasis or Patients on medication that influence bone metabolism as: glucocorticoid, heparin, anticonvulsant, thyroxin, hormone replacement therapy or any drug used in treatment of osteoporosis or previously received any anti TNFα therapy were excluded from the study.

Thorough clinical assessment

All patients were subjected to the following before starting treatment and 1 year after, full medical history taking with special emphasis on: age, sex, disease duration, morning stiffness duration, global pain assessment using Visual Analog Scale (VAS, 0-10) cm, modified Health Assessment Questionnaire (MHAQ) Disability Index [26] and type of drug therapy. Tenderness count (TJC), swollen joint count (SJC) and Extra- articular manifestations (EAM).

Complete blood count (CBC) was done including haemoglobin level. Erythrocyte sedimentation rate (ESR) by the Westergen method. C−Reactive Protein (CRP) level by nephelometry and rheumatoid factor (RF) by qualitative method. The 28-joint count Disease Activity Score (DAS28) was calculated using (CRP) level [27]. Disease activity status of remission (REM), low disease activity (LDA), moderate disease activity (MDA), and high disease activity (HDA) were also determined using DAS28. REM was defined as DAS 28 < 2.6, LDA as 2.6 ≤ DAS28 ≤ 3.2, MDA as 3.2 < DAS28 < 5.1, HDA as 5.1 ≤ DAS28 [28].

Radiological investigations

Plain X−ray: P−A view of both hands and wrists were assessed using the Van Der Heide Sharp score (SHS) for the extent of joint damage, as indicated by joint space narrowing and erosion. Sixteen joint areas and 16 joint areas in each hand were scored for erosion and joint space narrowing, respectively. The total Sharp Score (TSS) was calculated as the sum of the erosion and joint space narrowing scores [29]. ∆SHS ≥ 1 unit/year was regarded as radiographic progression according to the previously used definition [30].

Dual-energy X−ray absorptiometry (DEXA): By using GE Medical Systems, LUNAR (DPX–MD+) device. Bone mineral density (BMD) was measured before and after treatment at 3 sites which are lumbar spine, femoral neck, and distal radius. World Health Organization (WHO) diagnosis T score criteria were applied to BMD measurement [31].

Measurement of serum IL−34 and Cathepsin k levels: Serum samples were obtained from 30 patients with RA before starting treatment and 1 year after. Serum samples were stored at −80°C until analysis. Interleukin−34 (IL−34) and Cathepsin K(Cath-K) have been estimated by using sandwich enzyme immunoassay (ELISA) technique (WKEAMedSupplies, Changchun,China) as supplied with kit from WKEAMedSupplies Company, China (tom.aoke@hotmail.com). All biochemical measures were performed in a single batch and a comparable number of patient and control samples were always assayed simultaneously in the same ELISA plate. Cut off points in ng/L were calculated from our control group.

Statistical analysis

IBM SPSS statistics (V. 22.0, IBM Corp., USA, 2013) was used.
for data analysis. Data were expressed as Mean ± SD for quantitative parametric measures in addition to Median and Percentiles for quantitative non-parametric measures.

The following tests were done:
1. Comparison between two independent groups for non-parametric data using Wilcoxon Rank Sum test.
2. Comparison between more than 2 patient groups for non-parametric data using Kruskall Wallis test.
3. Ranked Spearman correlation test to study the possible association between each two variables among each group for non-parametric data.
4. Diagnostic sensitivity, specificity, predictive values, efficacy and accuracy by diagnostic validity test.
5. Logistic Multi-Regression analysis was used to search for a panel (independent parameters) that can predict the dependent variable parameter. The probability of error at 0.05 was considered sig., while at 0.01 and 0.001 are highly significant.

Results

The study included 41 RA patients enrolled from Physical Medicine, Rheumatology and Rehabilitation Department, Ain Shams University. They were randomly classified according to type of treatment into: group of patients included 21 RA patients: 17 females (80.95%) and 6 males (33.33%) treated with DMARDs (methotrexate (n=12), hydroxyl chloroquine (n=10), leflunomide (n=8) and low dose (5 mg) Prednisone (n=11). The comparison revealed that one year post treatment, patients had highly significant lower ESR, TJC and SJC (p<0.001) than baseline change.


<table>
<thead>
<tr>
<th>Data</th>
<th>Pre ttt</th>
<th>Post ttt</th>
<th>Z</th>
<th>p</th>
<th>Sig.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age(years)</td>
<td>45 (39−50)</td>
<td>39 (33.5−41)</td>
<td></td>
<td></td>
<td>NS</td>
</tr>
<tr>
<td>Disease Duration(years)</td>
<td>13 (10.5−20)</td>
<td>3 (2.5−5)</td>
<td></td>
<td></td>
<td>S</td>
</tr>
<tr>
<td>Morning stiffness (hr)</td>
<td>2 (1.5−2.25)</td>
<td>2 (1.5−2.5)</td>
<td></td>
<td></td>
<td>NS</td>
</tr>
<tr>
<td>Pain by VAS(cm)</td>
<td>9.5 (9−10)</td>
<td>9 (9−9.5)</td>
<td></td>
<td></td>
<td>S</td>
</tr>
<tr>
<td>MHAQ</td>
<td>1.77 (1.9−2.25)</td>
<td>1.8 (1.125−1.925)</td>
<td></td>
<td></td>
<td>NS</td>
</tr>
<tr>
<td>ESR(mm)</td>
<td>50 (45−54)</td>
<td>45 (40−52)</td>
<td></td>
<td></td>
<td>S</td>
</tr>
<tr>
<td>CRP(mg/dl)</td>
<td>30 (18−50)</td>
<td>44 (32−48)</td>
<td></td>
<td></td>
<td>NS</td>
</tr>
<tr>
<td>Hb level (g/dl)</td>
<td>11.5 (10.55−12)</td>
<td>11.3 (10.95−12.35)</td>
<td></td>
<td></td>
<td>S</td>
</tr>
<tr>
<td>TJC</td>
<td>20 (18−26)</td>
<td>16 (16−18)</td>
<td></td>
<td></td>
<td>S</td>
</tr>
<tr>
<td>SJC</td>
<td>10 (8−13)</td>
<td>10 (8−12)</td>
<td></td>
<td></td>
<td>NS</td>
</tr>
<tr>
<td>DAS28 –CRP</td>
<td>6.54 (5.925−6.73)</td>
<td>5.22 (4.81−5.345)</td>
<td></td>
<td></td>
<td>NS</td>
</tr>
<tr>
<td>DEXA T score</td>
<td>-2.8 (-3.5−1.9)</td>
<td>-1.9 (-2.25−1.5)</td>
<td></td>
<td></td>
<td>NS</td>
</tr>
<tr>
<td>SHS (0-314)</td>
<td>87 (62.5−115)</td>
<td>66 (54.5−79.5)</td>
<td></td>
<td></td>
<td>NS</td>
</tr>
<tr>
<td>IL-34 level (ng/L)</td>
<td>950 (740−1250)</td>
<td>125 (125−375)</td>
<td></td>
<td></td>
<td>NS</td>
</tr>
<tr>
<td>Cathepsin K (ng/L)</td>
<td>5.5 (3.75−6.625)</td>
<td>1.5 (0.75−1.75)</td>
<td></td>
<td></td>
<td>NS</td>
</tr>
</tbody>
</table>

Table 2: Comparison of patients characteristics before and after anti TNF α therapy. Pre ttt: pre treatment. Post ttt: post treatment patients group data. Pain, CRP, DAS28, DEXA scan. IL34, cathepsin K were significantly decreased post treatment.
significant difference in ∆ MS, ∆pain, ∆SJC, ∆DAS 28, ∆DEXA score, ∆SHS, ∆IL34 and ∆ cathepsin K between patients and controls.

Table 3: Comparison between patients and controls regarding change in variables (∆) before and after treatment. ∆MS: change in morning stiffness. This table showed a significant difference in ∆ MS, ∆pain, ∆SJC, ∆DAS 28, ∆DEXA score, ∆SHS, ∆IL34 and ∆ cathepsin K between patients and controls.

<table>
<thead>
<tr>
<th>Variable</th>
<th>Patients</th>
<th>Controls</th>
<th>Z</th>
<th>P</th>
<th>Sig</th>
</tr>
</thead>
<tbody>
<tr>
<td>∆ MS</td>
<td>-0.5 (-0.667−0.354)</td>
<td>0 (-0.225−0)</td>
<td>-3.362</td>
<td>0.001</td>
<td>HS</td>
</tr>
<tr>
<td>∆ Pain</td>
<td>-0.3 (-0.528−0.211)</td>
<td>-0.222 (-0.236−0.111)</td>
<td>-2.282</td>
<td>0.022</td>
<td>S</td>
</tr>
<tr>
<td>∆ MHAQ</td>
<td>-0.111 (-0.355−0.037)</td>
<td>-0.189 (-0.245−0.112)</td>
<td>-0.385</td>
<td>0.7</td>
<td>NS</td>
</tr>
<tr>
<td>∆ ESR</td>
<td>-0.378 (-0.481−0.245)</td>
<td>-0.25 (-0.315−0.213)</td>
<td>-1.91</td>
<td>0.056</td>
<td>NS</td>
</tr>
<tr>
<td>∆ CRP</td>
<td>-0.556 (-0.75−0.417)</td>
<td>-0.5 (-0.531−0.188)</td>
<td>-1.548</td>
<td>0.122</td>
<td>NS</td>
</tr>
<tr>
<td>∆ Hb level</td>
<td>0.103 (0.009−0.131)</td>
<td>0.028 (-0.013−0.047)</td>
<td>-1.7</td>
<td>0.089</td>
<td>NS</td>
</tr>
<tr>
<td>∆ TJC</td>
<td>-0.444 (-0.538−0.3)</td>
<td>-0.333 (-0.354−0.25)</td>
<td>-1.482</td>
<td>0.138</td>
<td>NS</td>
</tr>
<tr>
<td>∆ SJC</td>
<td>-0.438 (-0.75−0.333)</td>
<td>-0.25 (-0.388−0.208)</td>
<td>-2.749</td>
<td>0.006</td>
<td>HS</td>
</tr>
<tr>
<td>∆ DAS28</td>
<td>-0.253 (-0.268−0.157)</td>
<td>-0.121 (-0.193−0.068)</td>
<td>-2.196</td>
<td>0.028</td>
<td>S</td>
</tr>
<tr>
<td>∆ DEXA score</td>
<td>-0.125 (-0.27−0)</td>
<td>0.133 (0.053−0.23)</td>
<td>-3.94</td>
<td>0</td>
<td>HS</td>
</tr>
<tr>
<td>∆ SHS</td>
<td>0.067 (0−0.152)</td>
<td>0.159 (0.107−0.17)</td>
<td>-1.977</td>
<td>0.048</td>
<td>S</td>
</tr>
<tr>
<td>∆IL-34 (ng/L)</td>
<td>-0.315 (-0.355−0)</td>
<td>0.333 (0−0.4)</td>
<td>-3.248</td>
<td>0.001</td>
<td>HS</td>
</tr>
<tr>
<td>∆ Cathepsin K (ng/L)</td>
<td>-0.25 (-0.354−0.033)</td>
<td>0.167 (0−0.304)</td>
<td>-3.212</td>
<td>0.001</td>
<td>HS</td>
</tr>
</tbody>
</table>

Table 4: Diagnostic Validity Test of IL-34 for highly active R. PPV: positive predictive value, NPV: negative predictive value.

<table>
<thead>
<tr>
<th>Cut off value</th>
<th>Test of accuracy</th>
<th>Sensitivity</th>
<th>Specificity</th>
<th>PPV</th>
<th>NPV</th>
</tr>
</thead>
<tbody>
<tr>
<td>IL-34 At titre 375 ng/L</td>
<td>96.7 %</td>
<td>95.2 %</td>
<td>100.0 %</td>
<td>100 %</td>
<td>90 %</td>
</tr>
</tbody>
</table>

Table 5: Diagnostic Validity Test of cathepsin K for RA with osteoporosis.

<table>
<thead>
<tr>
<th>Cut off value</th>
<th>Test of accuracy</th>
<th>Sensitivity</th>
<th>Specificity</th>
<th>PPV</th>
<th>NPV</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cathepsin K At titre 2 ng/L</td>
<td>90 %</td>
<td>90 %</td>
<td>88.9 %</td>
<td>95.0 %</td>
<td>80 %</td>
</tr>
</tbody>
</table>

(Tables 4). The most sensitive and specific cut off value for cathepsin K as a diagnostic test of RA with osteoporosis is 2ng/L with accuracy of 90% (Table 5). Further analysis of our results to find any association of serum IL-34 with patients’ characteristics revealed that baseline IL-34 was positively correlated with baseline disease duration, ESR, CRP and MHAQ (r = 0.852, 0.714, 0.432, 0.685 P< 0.05) respectively and with SJC, DAS28 and SHS (r = 0.515, 0.490, 0.421, P< 0.05) respectively after treatment. Also the change in serum IL-34 after treatment was negatively correlated with DEXA T scores (r = -0.433, p< 0.05).

Similarly, Correlation of baseline serum Cathepsin K with other parameters revealed a positive correlation with SJC, DAS28, SHS and DEXAT scores (r = 0.774, 0.503, 0.711, 0.892, P< 0.05) respectively.

Relation of IL34 cytokine to cathepsin K enzyme

Correlation of ∆ IL–34 with ∆ Cathepsin K: There was a statistical significant positive correlation (r = 0.523, P< 0.05) between change in serum IL–34 level and change in serum Cathepsin K level throughout the study.

Response according to WHO/ILAR response criteria of improvement

Further analysis of the results revealed that 15 patients were improved (good responders) according to WHO/ILAR response criteria of improvement [53]:

1. >20% improvement in swollen joint count
2. >20% improvement in tender joint count, or > 5 if the count is between 16 and 20
3. >20% improvement in at least two of the following three measures: (i) patient's or physician's global disease activity; (ii) pain; (iii) erythrocyte sedimentation rate. While 6 patients weren’t (poor responders) at the end of the study. Comparison between the 2 subgroups regarding clinical, laboratories and radiological variables was done (Table 6).

Relation of IL-34 and cathepsin k to DEXA grading

Kruskal Wallis test was used to compare serum IL-34 levels after treatment among the three DEXA subgroups (osteopenic, osteoporotic, severe osteoporotic) and revealed a significant difference between the 3 subgroups (H= 8.795, p = 0.012) as well as significant difference between osteopenic and osteoporotic grades (Z=−2.12, p = 0.034).

Similarly comparison of serum cathepsin K levels after treatment among all grades of DEXA was studied and revealed high significant difference between the 3 grades (H =14.362, p=0.002) (Table 7).

The target of this study was to measure serum levels of IL-34 and Cathepsin K in active long standing RA patients before and after treatment with anti TNF α drugs vs. early RA patients on conventional DMARDS and to evaluate the effect of anti TNF α drugs on disease progression (joint destruction and osteoporosis) in RA patients, so effect of biologic therapy on radiological progression (∆ SHS/year) was measured across treatment time. After one year period of follow up the median (∆ SHS/year) was 0.067 (0−0.152) which is less than one unit that considered no progression according to previously used definition.

Predictors of radiologic change

To search for predictors that can predict the less radiographic change, by using logistic stepwise multi-regression analysis in 3 models, we get the most sensitive ones which was high baseline DAS28 and using TNF α inhibitors (Table 8).

Third model showed that high baseline DAS28 and using TNF α inhibitors were the most sensitive predictors for radiologic change.
Comparison between good and bad responders. This table showed a significant difference in morning stiffness, SJC, DAS 28, and serum IL-34 in good responders versus poor responders.

<table>
<thead>
<tr>
<th>Variable</th>
<th>Good Responders Median(25-75)</th>
<th>Poor Responders Median(25-75)</th>
<th>Z</th>
<th>P</th>
<th>Sig</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age</td>
<td>42(39-50)</td>
<td>45.5(44.5-53.75)</td>
<td>-1.33</td>
<td>0.183</td>
<td>NS</td>
</tr>
<tr>
<td>Disease duration</td>
<td>13(10-23)</td>
<td>13.5(11.75-20)</td>
<td>-0.431</td>
<td>0.686</td>
<td>NS</td>
</tr>
<tr>
<td>MS*</td>
<td>0.75(0.5-1)</td>
<td>2(1.25-5.625)</td>
<td>-2.678</td>
<td>0.007</td>
<td>HS</td>
</tr>
<tr>
<td>Pain(VAS)</td>
<td>6(4-7)</td>
<td>7(6.375-8.25)</td>
<td>-1.424</td>
<td>0.155</td>
<td>NS</td>
</tr>
<tr>
<td>TJC</td>
<td>12(8-14)</td>
<td>14(10-20)</td>
<td>-1.504</td>
<td>0.133</td>
<td>NS</td>
</tr>
<tr>
<td>SJG*</td>
<td>4(1-8)</td>
<td>8(7.5-9)</td>
<td>-2.315</td>
<td>0.021</td>
<td>S</td>
</tr>
<tr>
<td>CRP*</td>
<td>12(6-12)</td>
<td>18(10.5-28.5)</td>
<td>-2.132</td>
<td>0.043</td>
<td>S</td>
</tr>
<tr>
<td>DAS 28*</td>
<td>4.7(4.21-4.98)</td>
<td>5.35(4.8975-6.14)</td>
<td>-2.315</td>
<td>0.021</td>
<td>S</td>
</tr>
<tr>
<td>SHS</td>
<td>90 (68-124)</td>
<td>89.5(71.25-165)</td>
<td>-0.624</td>
<td>0.533</td>
<td>NS</td>
</tr>
<tr>
<td>DEXA</td>
<td>-2.5(-3.4-1.2)</td>
<td>-2.65(-3.75-1.817)</td>
<td>-0.546</td>
<td>0.585</td>
<td>NS</td>
</tr>
<tr>
<td>HAQ(DI)</td>
<td>1.25(1-2.12)</td>
<td>2.05(0.89-2.25)</td>
<td>-0.665</td>
<td>0.506</td>
<td>NS</td>
</tr>
<tr>
<td>Cathepsin K</td>
<td>3.75(3-5.5)</td>
<td>5.25(3.18-10.12)</td>
<td>-1.133</td>
<td>0.257</td>
<td>NS</td>
</tr>
<tr>
<td>IL-34*</td>
<td>520(500-770)</td>
<td>937(760-1187.5)</td>
<td>-2.787</td>
<td>0.005</td>
<td>HS</td>
</tr>
</tbody>
</table>

Table 6: Comparison between good and bad responders. This table showed a significant difference in morning stiffness, SJC, DAS 28, and serum IL-34 in good responders versus poor responders.

<table>
<thead>
<tr>
<th>Grade</th>
<th>Z</th>
<th>P</th>
<th>Sig</th>
</tr>
</thead>
<tbody>
<tr>
<td>Grade 1</td>
<td>-4.02</td>
<td>0.002</td>
<td>HS</td>
</tr>
<tr>
<td>Grade 2</td>
<td>14.362</td>
<td>0.002</td>
<td>HS</td>
</tr>
</tbody>
</table>

Table 7: Comparison of serum Cathepsin K among DEXA grades. This table showed a high significant difference between the 3 grades of DEXA.

Table 8: Stepwise multiple linear regression analysis for ∆SHS. These 3 Models showed that high baseline DAS28 and using TNF α inhibitors were the most sensitive predictors for ∆ DEXA and F ratio was 3.954, p=0.038.

Stepwise multiple linear regression analysis for ∆ DEXA was done and showed that high baseline CRP and using TNF α inhibitors were the most sensitive predictors for ∆ DEXA and F ratio was 3.954, p=0.038.

Discussion

Rheumatoid arthritis is characterized by subchondral bone loss and the mechanism of bone loss is similar or equal to that of osteoporosis [32]. High BMD loss in RA patients was associated with joint damage progression, disease activity, functional disability and immobility in previous longitudinal studies, even in early RA [33]. Interleukin 34 (IL-34), a recently discovered cytokine that binds macrophage colony-stimulating factor (M-CSF) receptor [34]. Several recent studies revealed that IL-34 expression increases in the synovium, sera, and SFs from patients with RA as well as an association between IL-34 levels and RF and anticytic citrullinated peptide (CCP) antibody titers [11,34]. It has been known that CSF-1 is present in synovial fibroblasts, plasma, and SF from patients with RA, and that CSF-1R is upregulated in RA synovium [15]. In addition, the administration of anti-CSF-1R antibody reduces the severity of collagen induced arthritis (CIA). After all, it was considered that IL-34 could play a significant role in the synovial inflammation of RA [35]. Functionally, isolated RA-derived fibroblast-like synoviocytes and osteoblasts were found to produce IL-34 in response to TNFa [36]. It was reported that anti TNF-a induced down-regulation of membrane RANKL could be important in preventing articular damage in RA patients. Joint damage may also be mediated by other mediators involved in osteoclast functions such as cathepsin-K [37]. Moreover, the osteoclastogenesis factor RANKL, appears to directly up-regulate cathepsin K expression [38]. Interestingly In the synovium of RA, the cathepsin K protein was localized in synovial fibroblasts, stromal multinucleated giant cells and CD68+ macrophage-like synoviocytes. The overexpression of cathepsin K in RA synovia proves that this protease may become a new and highly specific biomarker for RA.

As previous studies mentioned the pro-osteoclastogenic role of IL-34 and its response to pro-inflammatory cytokines such as tumor necrosis factor-α [39]. Accordingly we suggested that IL-34 can be down regulated under anti TNF-a treatment in human RA, especially in the aspect of joint destruction.

To our knowledge this is the first study for investigating serum levels of both IL34 and cathepsin K as inflammatory and osteoclastogenic factors in early and established RA, before and after anti TNFa therapy versus conventional DMARDS in relation to localized (Joint destruction) and generalized osteoporosis.

We found a significant higher serum IL-34 and Cathepsin K level in patients with established RA who will receive a biologic therapy than in those with early RA (controls). This could be explained by the difference in disease stage and severity as there was a significant increase in age, disease duration, number of tender joint count, DEXA scores more in patients group than in controls.

Batmaz et al. found that cathepsin K levels were significantly higher in patients with post menopause RA when compared with that of the post menopause healthy control group (p<0.05). In addition, the elevated serum levels of cathepsin K were positively correlated with disease activity score (DAS 28) and total Larsen scores (hands, feet and total) (p<0.05) [40]. In addition, Skoumal et al., noticed that cathepsin K serum levels of the patients with RA were significantly elevated (P=0.0003) compared with the healthy control group and a statistically significant correlation between cathepsin K and the Larsen score (P=0.004) [41].

The effect of anti-TNFα therapy was clear by the significant decrease in symptoms and signs of synovitis (pain, TJC, SJC) and activity (ESR, CRP, DAS28-CRP) along with s. IL-34 and s.cathepsin K, and improvement in DEXA T scores at the end of the study in patients group while the reduction in s.IL-34 and s.cathepsin-K, and DAS28 didn’t reach significant level (p>0.05) in controls group and DEXA scores were deteriorated. This was in accordance with Tian et al, who reported that the level of serum IL-34 decreased after anti-TNF treatment in RA patients [14]. TNF inhibitors delay appearance of bone erosion in RA...
patients with no progression of bone destruction in responders and possibly to some extent, in non-responders [42] and Prevention of bone loss [43,44].

On the other hand, our findings of reduction in cathepsin k level post treatment was in contrary with Cauli et al., who reported a persistent increase in cathepsin-K in RA patients under adalimumab (ADA) up to 24 weeks [37]. This may be due to shorter duration of treatment (6 months) than our duration (12 months) and trying two TNFα inhibitors drugs working by two different mechanisms of action: Etanercept (as a soluble receptor antagonist) and ADA (as a monoclonal antibody to TNF-α), therefore, this could successfully decrease cathepsin k level along the study time, reduce and systemic osteoporosis as evident by DEXA T scores were improved in patients post biologic therapy. Moreover, Cauli et al., reported that the inhibition of joint damage and osteoporosis seen in patients treated with anti-TNFα drugs could be due to reduction in other contributors (DKK-1, sclerostin) in RA rather than reduction in cathepsin k [37]. Moreover, other MMPs and cathepsins were known as potential contributors to bone destruction [45].

Furthermore, although most of patients were improved under biologic therapy, there was no significant difference between patients received Etanercept vs. those received ADA, this may be due to the equal efficacy of both drugs. Our findings are similar to Scott and Kingsley, stated that the TNF inhibitors have exhibited a superior ability to reduce the signs and symptoms of RA, inhibit progression of structural damage and improve physical function in patients with this disease. But the question as to which of the TNF inhibitors affords the greatest efficacy cannot be answered and the choice of agent therefore depends on other factors, including patients’ convenience, access to treatment and cost [46].

In our study, to elaborate the effect of anti TNFα versus DMARDs it was obvious that change in radiologic joint progression per year (ASHS) was significantly less in patients than in controls, moreover, a significant difference in DEXA-T scores was noticed reflecting improvement of osteoporosis in patients [-0.125 (-0.27-0)] while deterioration in controls [0.133 (0.053-0.23)] although baseline T scores were higher in patients versus controls. Additionally, Δ IL-34 and Δ cathepsin k were higher in patients under anti TNFα therapy than in controls under conventional DMARDs (Table 3). Vis et al., found that no bone loss was observed in the spine and hip, while a decrease of BMD was observed at the hands after 1 year of infliximab therapy in 102 RA patients [47]. This was in agreement with Marotte et al., who found a significant decrease of BMD (-3.4% at the femoral neck and -3.9% at lumbar spine, p < 0.001) in active RA patients treated with methotrexate alone while no decrease was observed in the group treated by infliximab and methotrexate [43].

Marotte and Miossec, concluded that TNFα blockade is not only able to prevent joint destruction, but it is also able to prevent bone loss in RA patients [48].

It was clear that IL-34 is strongly associated with disease activity and radiologic progression as IL-34 was positively correlated with morning stiffness, SHS, ESR and CRP (p< 0.05). This was in accordance with a recent study by Chang et al. that baseline IL-34 levels were positively correlated to ASHS/year [49]. As it was previously confirmed that SHS was positively correlated with disease activity and local inflammation and this was similar to our findings [50]. Similarly, we found that serum cathepsin k before and after treatment was strongly correlated with TJC, SJC, DAS28, SHS and DEXA T scores which confirms the imperative role of cathepsin k in pathogenesis of RA in both synovitis and joint destruction as well as generalized osteoporosis. This was in accordance with Skoumal et al., proved that cathepsin K is upregulated in the serum and synovial fluid of RA patients with bone degradation. Moreover, its serum concentrations significantly correlated with radiological joint destruction in RA patients [41].

Moreover, a positive correlation (P<0.05) between change in serum IL–34 level and change in serum Cathepsin K level throughout the study; and both of them were significantly correlated with DEXA T scores (p<0.05), strongly reflects the involvement of both novel markers in disease activity and severity as well as the intimate link between cathepsin k and IL-34 as both of two mediators expressed and exerted joint destruction and osteoporosis in RA through RANKL [38,10]. Moroko et al., mentioned that overexpression of cathepsin K leads to spontaneous synovitis and cartilage erosion in RA [51].

After classification the patients under anti TNFα therapy into good and poor responders subgroups, we found significant reduction of morning stiffness, SJC, CRP, DAS28 and serum IL-34 in good responders versus poor responders (p<0.01). Similarly further reduction in serum cathepsin k was found in good responders [3.75(3.5-5.5)] versus poor responders [5.25(3.18-10.12)] which reflected that impact of treatment response was variable on patients of the same group. It may be due to other contributing poor factors as RF positivity, age, gender and disease duration, as three of them were male and all were seropositive and their disease duration range (12-20) years which declare that pathogenesis of RA may differ from a patient to another (disease severity), as well as effect of treatment varied also from one to another patient and this explanation was previously established by many authors [52].

Consequently, in our study, an obvious association was obvious between circulating IL-34 and generalized osteoporosis as a significant difference existed post treatment between s.IL-34 and the three grades of DEXA T scores (H= 8.795, p = 0.012) and even between any two grades of DEXA T scores. This indicated the potential role of IL-34 in osteoactogenesis. However, this was in contrary to Moon et al., who reported that no significant correlation was found between serum IL-34 and systemic osteoporosis although IL-34 concentration in synovial fluid was significantly correlated with RANKL levels in his study [34]. On the other hand, our findings for such correlation could be explained by the theory that IL-34 promotes the formation of macrophage colonies from human bone marrow equivalently to CSF-1 and was found to be able to substitute for CSF-1 in receptor activator of nuclear factor kappa-B ligand (RANKL)-induced osteoactogenesis 3 and 4 [10].

Similarly, when comparing serum level of cathepsin k between subgroups of DEXA T scores, a significant difference existed between them and even between grades of T scores, which reflects that cathepsin k is highly relevant to osteoporosis, with intimate relation to minimal change in bone mineral density, this was clear in our study as cathepsin k had a 90% sensitivity and accuracy to detect RA with osteoporosis at a cutoff value 2 ng/L. Whereas some of the non-osteoclast cell types exhibiting cathepsin K expression open a window of opportunity to consider cathepsin K as a novel target for other disease (chondrocytes: osteoarthritis; synovial fibroblasts: rheumatoid arthritis; macrophages/giant multinucleated cells: atherosclerosis), other sites of expression may raise concern [32].
Conclusion

We found that serum IL-34 and cathepsin k were upregulated in peripheral blood of longstanding RA versus early RA patients. Both were strongly linked to disease activity and were highly relevant to both localized osteoporosis in the form of joint destruction and generalized osteoporosis in the form of DEXA T scores. Furthermore, anti TNF-α therapy effectively decreased both biomarkers regardless drug type, with amelioration of clinical, laboratory and radiological parameters of RA patients.

Acknowledgement

All authors declare that there was no conflict of interest, no funding sources in preparing this work.

References

expressed in gingival fibroblasts, shows enhanced expression by pro-inflammatory cytokines, and stimulates osteoclast differentiation. PloS One 8: e81665.


