Alpha amylase and alpha glucosidase inhibitory activities of isolated total saponins and tannins of Dialium guineense stem bark

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Abstract

Background: Dialium guineense belongs to the Leguminosae family, and grows in dense forests in Africa along the southern edge of the Sahel. The bark, leaves and fruits of the plant have medicinal properties and are used to treat diseases such as stomatitis, toothache, fever, diarrhoea, palpitations, and microbial infections.

Aim: The present study investigated the α -amylase and α -glucosidase inhibitory activities of isolated total saponins and tannins of Dialium guineense stem bark.

Methods: Total saponins and tannins were isolated from D. guineense stem bark using standard method. Alpha amylase and alpha glucosidase inhibitory activities of the extracts were determined in vitro. Acarbose was used as the reference drug.

Results: The results showed that both extracts of Dialium guineense possess α -amylase and α -glucosidase inhibitory activities. The total tannins showed better α -amylase and α -glucosidase inhibitory activities when compared with the total saponins extract (p < 0.05). The enzyme activities of the extracts were concentration-dependent, and comparable to those of acarbose.

Conclusion: The results of this study indicate that Dialium guineense shows great potential in the treatment of Diabetes mellitus.

Keywords: Alpha amylase, Alpha glucosidase, Dialium guineense, Acarbose, Inhibitory activity.

Introduction

Medicinal plants contain wide varieties of chemical substances which possess important therapeutic properties that can be utilized in the treatment of human diseases (Gowthami et al., 2012a; Gowthami et al., 2012b; Manokaran et al., 2008).

Dialium guineense (Velvet Tamarind), is a tall, tropical, fruit-bearing tree. It belongs to the Leguminosae family, and has small, typically grape-sized edible fruits with brown hard inedible shells. In Africa, it grows in dense forests along the southern edge of the Sahel. The bark and leaves have been reported to possess medicinal properties and are used against several diseases. Each fruit typically has one hard, flat, round, brown seed, typically 7 - 8 mm across and 3 mm thick (Dalziel and Hutchison, 1973). The seed somewhat resembles a watermelon seed (Citrullus lanatus). Some have two seeds. The seeds are shiny, and coated with a thin layer of starch. The pulp is edible and may be eaten raw or soaked in water and consumed as a beverage (Dalziel and Hutchison, 1973). The bitter leaves are ingredients in a Ghanaian dish called domoda. Its wood is hard and heavy, and used for construction. The wood is also used for firewood and charcoal production (Dalziel and Hutchison, 1973).

Saponins are a class of chemical compounds found in abundance in various plants species. They are amphipathic glycosides grouped phenomenologically by the soap-like foam they produce in aqueous solutions when shaken, and structurally by having one or more hydrophilic glycoside moieties combined with a lipophilic triterpene or steroid derivative (Hostettmann and Marston, 1995).

Tannins are phenolic compounds of high molecular weight, which are soluble in water and alcohol, and are found in the root, bark, stem and outer layers of plant tissue. Their acidic nature in reactions is attributed to the presence of phenolic or carboxylic acid group (Kar, 2007).

Before the emergence of oral hypoglycemic agents, the major form of treatment for diabetes mellitus involved dietary manipulation and the use of plant therapies. About 400 plants world-wide have been documented for the treatment of diabetes mellitus and the majority awaits proper scientific and medicinal evaluation. Studies have shown that the antidiabetic mechanisms of traditional herbs include reduced carbohydrate absorption (De Fronzo, 2004), reduced α-glucosidase, α-amylase, and aldose reductase activities (Velho and Froguel, 1997), increased glucose uptake in muscle and adipose tissues (Wang et al., 2013), activation of PPAR y (Kandra, 2003), increased insulin sensitivity/upregulation of receptor expression (Dewi et al., 2007), exertion of antioxidant effects and decreased β-cell apoptosis (Cheng and Fantus, 2005), stimulation of β-cell insulin secretion (Marles and Farnsworth, 1995), inhibition of hepatic gluconeogenesis/glycogenolysis (De Sales, 2012), and prevention of endogenous incretins from degradation/ suppression of glucagon (Wang, 2013).

Pancreatic α -amylase, a calcium metalloenzyme that catalyzes the hydrolysis of α -1, 4 glycosidic linkages of amylose, amylopectin, glycogen, and various maltodextrins, is responsible for most of starch digestion in humans. A positive correlation between human pancreatic α -amylase (HPA) activity and increase in postprandial glucose levels has been shown, demonstrating the relevance of suppressing postprandial hyperglycemia (PPHG) in the treatment of type 2 diabetes mellitus (T2DM) (Karthic et al., 2008).

Alpha glucosidase is an enzyme which is responsible for the conversion of complex carbohydrates into glucose. This enzyme acts by delaying the breakdown of complex carbohydrates into glucose and reduces its absorption from the gut which results in reduction of postprandial blood sugar level (Subramanian et al., 2008). Although commercially available α -glucosidase inhibitors like voglibose and acarbose are used effectively to control blood glucose levels, they have been associated with serious gastrointestinal side effects. Therefore, there is a need to search for an alternative that can exhibit α -glucosidase inhibitory activity without side effects (Playford et al., 2013). The present study investigated the α -amylase and α -glucosidase inhibitory activities of extracts of Dialium guineense stem bark.

MATERIALS AND METHODS

Plant Sample Collection and Preparation

The plant leaves were obtained from lyekogba area in Benin and identified at the Department of Plant Biology and Biotechnology, University of Benin, Benin City, Nigeria with voucher number UBHD330, after which the bark was obtained. Preparation and

isolation was carried out using the methods of Hostettmann and Marston, (1995) and Haslam, (1996).

Determination of D. guineense Alpha-amylase Inhibitory Activity

The assay mixture containing 200 μL of 0.02 M sodium phosphate buffer, 20 μL of enzyme and the plant extracts (20 - 100 $\mu g/mL$) were incubated for 10 min at room temperature followed by addition of 200 μL of starch to all test tubes. The reaction was terminated with the addition of 400 μL dinitrosalicylate (DNS) reagent and placed in boiling water bath for 5 min, cooled and diluted with 15 mL of distilled water and absorbance was read at 540 nm. The control samples were prepared without any plant extracts. The % inhibition was calculated according to Equation 1:

Inhibition (%) = A control – Aextract
$$x$$
 100.....(1)

Acontrol

where A is absorbance.

The half maximum inhibitory concentration (IC50) values were determined from plots of percentage inhibition versus log of inhibitory concentration and were calculated using nonlinear regression analysis from the mean inhibitory values. Acarbose was used as the reference alpha amylase inhibitor. All tests were performed in triplicate (Jung et al., 2006).

Determination of D. guineense Alpha-glucosidase Inhibitory Activity

The yeast alpha glucosidase used was dissolved in 100 mM phosphate buffer pH 6.8. P-Nitrophenyl- α -D-glucopyranoside was used as the substrate. Plant extracts (20 - 100 μ g/mL) were mixed with 320 μ L of 100 mM phosphate buffer pH 6.8 at 30 °C for 5 min. Then, 3 mL of 50 mM NaOH was added to the mixture and the absorbance was read at 410 nm. The control samples were prepared without any plant extracts. The % inhibition was calculated according to Equation 2:

Inhibition(%)=Acontrol-Aextractx100......(2)

Acontrol

The IC50 values were determined from plots of percentage inhibition versus log of inhibitory concentration and were calculated using nonlinear regression analysis from the mean inhibitory values. Acarbose was used as the reference alpha glucosidase inhibitor. All tests were performed in triplicate (Jung et al., 2006).

Statistical Analysis

Data are presented as mean \pm SEM. Statistical analysis was performed using SPSS (21.0). Statistical significance was assumed at p < 0.05.

RESULTS

Alpha-amylase and Alpha-glucosidase Inhibitory Activities of Isolated Total Saponins and Tannins of D. guineense Stem Bark

The results showed that both extracts of Dialium guineense possess α -amylase and α -glucosidase inhibitory activities. The total tannins showed better α -amylase and α -glucosidase inhibitory activities when compared with the total saponins extract (p < 0.05). The enzyme activities of the extracts were concentration-dependent, and comparable to those of acarbose. (the standard drug). These results are shown in Tables 1 and 2).

Concentration of drug or extract (pg/mL)	Percentage inhibition (%) of a amylase by extracts			
	Total Saponins	Total Tannins	Acarbove	
10	1.50 ± 0.09	3.00 ± 0.10	24.20 ± 1.20	
20	1.50 ± 0.08	4.50 ± 0.85	27.20 ± 1.08	
30	6.00 ± 0.10	10.60 = 0.71	30.30×1.50	
40	7.30 ± 0.35	25.70 ± 1.05	34.80 ± 1.22	
50	9.00 ± 0.70	39.50 ± 1.64	50.00 ± 1.58	
60	13.60 ± 1.00	45.40 ± 1.55	94.50 ± 1.55	
70	15.10 ± 0.85	51.50 ± 1.05	56.00 ± 1.05	
80	22.70 ± 1.02	66.60 ± 1.85	60.60 ± 1.10	
ICm (ng/mL)	185.98 ± 5.81	65.18 + 3.11	58.46 ± 3.08	

Data are percentage inhibition of $\alpha\text{-amylase}$ by extracts, and are expressed as mean \pm SEM

(n = 3).

Table 2: Percentage Inhibition of Alpha-glucosidase by Isolated Total Saponins and Tannins of Dialium guineense Stem Bark

Concentration of drug or extract (µg/mt)	Percentage inhibition (%) of a glucosidase by extracts Total Saponins Total Tannins Acarbose			
10	5.20 ± 0.10	1.40 ± 0.09	12.60 ± 0.64	
20	8.80 a 0.63	3.10 ± 0.11	14.00 ± 0.88	
30 40	11.20 ± 0.48	19.00 1 0.44	14.90 ± 0.85	
40	15.30 ± 0.84	20.00 ± 0.84	24.40 ± 1.41	
50	25.30 ± 1.21	20.70 ± 1.21	27.40 ± 1.09	
60	40.80 ± 1.44	45.40 ± 1.62	29.40 ± 1.64	
70	51.40 ± 1.82	51.40 ± 1.85	37.50 ± 1.65	
80	55.90 ± 1.05	63.20 ± 1.75	45.90 ± 1.95	
K to (mg/mt)	74.53 ± 2.88	69.48 ± 3.11	96.28 ± 3.10	

Data are percentage inhibition of $\alpha\text{-}\textsc{glucosidase}$ by extracts, and are expressed as mean \pm SEM

(n = 3).

DISCUSSION

Plant food rich in polyphenols have been reported to produce effects similar to insulin in the utilization of glucose, and act as good inhibitors of key enzymes associated with T2DM and lipid peroxidation in tissues (Reddy et al., 2010). Studies have shown that the bioactivity of polyphenols in plants is linked to their antioxidant properties and many of these plants also possess hypoglycemic effect (Ramkumar et al., 2010). Higher plants, animals and microorganisms produce a large number of different protein inhibitors of α -amylase and α -glucosidase in order to regulate the activity of these enzymes (Choudhury et al., 1996). Some of these inhibitors act by directly blocking the active site of the enzyme (Kavitha et al., 2012). In animals, α-amylase inhibitors decrease the high glucose levels that occurs after a meal by slowing the speed with which α-amylase converts starch to simple sugars (Boivin et al., 1987). This is important in diabetic patients where low insulin levels prevent rapid clearance of extracellular glucose from the blood (Mohammed et al., 2009). Hence diabetics tend to have reduced α-amylase activity in order to keep their glucose levels under control. Plants also use α-amylase inhibitors as defense mechanism against insects. These inhibitors alter the digestive action of α-amylase and proteinases in the gut of insects and inhibit their normal feeding behavior. Therefore, a-amylase inhibitors have potential roles in controlling blood sugar levels and crop protection (Kumanan et al., 2010). Alpha glucosidase inhibitors are used as oral antidiabetic drugs for treating T2DM. They act by preventing the digestion of carbohydrates such as starch. Alpha glucosidase inhibitors act as competitive inhibitors of a-glucosidase enzyme needed to digest carbohydrates. Intestinal α-glucosidase hydrolyzes complex carbohydrates to glucose and other monosaccharides in the small intestine. Inhibition of these enzyme systems helps to reduce the rate of digestion of carbohydrates (Bhat et al., 2011). Less amounts of glucose is absorbed because the carbohydrates are not completely broken down into glucose molecules. In diabetics, the short-term effect of these enzyme inhibitor drug therapies is to decrease high blood glucose levels. The presently used synthetic enzyme inhibitors cause gastrointestinal side effects such as diarrhea, flatulence, and abdominal bloating (Bray and Greenway, 1999). Therefore, natural α -amylase and α -glucosidase inhibitors from dietary plants can be used as an effective therapy for treating post-prandial hyperglycemia with minimal side effects. In this study, total saponins and tannins of Dialium guineense stem bark exhibited good in vitro α -amylase and α -glucosidase inhibitory activities which were comparable with those of acarbose the standard drug used.

CONCLUSION

The results of this study indicate that Dialium guineense shows great potential in the treatment of Diabetes mellitus.

REFERENCES

- Gowthami, M., Tamil-Selvi, S., Senthil-Kumar, G. and Panneerselvam, A. (2012a). Phytochemical analysis and antibacterial properties of leaf extract of Azima tetracantha (Lam.). Asian Journal of Plant Science and Research. 2 (2): 110 - 114.
- Gowthami, M., Tamil-Selvi, S., Senthil-Kumar, G. and Panneerselvam, A. (2012b). Phytochemical analysis and antibacterial properties of leaf extract of Azima tetracantha (Lam.). Asian Journal of Plant Science and Research. 2 (2): 89 – 94.
- Manokaran S., Jaswanth A., Sengottuvelu, S., Nandhakumar, J., Duraisamy, R., Karthikeyan, D. and Mallegaswari R (2008). Hepatoprotective Activity of Aerva lanata Linn. Against Paracetamol Induced Hepatotoxicity in Rats. Res J. Pharm Tech. 1 (4): 398 – 400.
- Dalziel, J.M. and Hutchison, J. (1973). Flora of West Tropical Africa. Vol.1 (2nd Ed). The White friars Press Ltd. London. Pp. 561.
- 5. Hostettmann, K. and Marston, A. (1995). Saponins. Cambridge: Cambridge University press. Pp. 3.
- Kar, A. (2007). Pharmacognosy and Pharmaco-biotechnology (Revised-Expanded Second Edition). New A g e International Limited Publishers, New Delhi. Pp. 332 - 600.
- De Fronzo, R.A. (2004). Pathogenesis of type 2 diabetes mellitus. Med Clin North America. 88 (4): 787 - 835.
- 8. Velho, G. and Froguel, P. (1997). Maturity-onset diabetes of the young (MODY), MODY genes and non-insulin-dependent diabetes mellitus. Diabetes Metabolism. 23 (2): 34 37.
- Wang, Z., Wang, J. and Chan, P. (2013). Treating Type 2 Diabetes Mellitus with Traditional Chinese and Indian Medicinal Herbs 17.
- 10. Kandra, L. (2003). α-Amylases of medical and industrial importance. J of Mol Structure. 666 (667): 487 498.
- 11. Dewi, R.T., Iskandar, Y.M., Hanafi, M., Kardono, L.B.S., Angelina, M., Dewijanti, I.D. and Banjarnahor, S.D.S. (2007) Inhibitory effect of Koji Aspergillus terreuson α-glucosidase activity and postprandial hyperglycemia. Pak J Biol Science. 10 (18): 3131 3135.
- 12. Cheng, A.Y.Y and Fantus I.G. (2005). Oral anti-hyperglycemic therapy for type 2 diabetes Mellitus. Can Med Assoc Journal. 172: 213 226.
- 13. Marles, R.J. and Farnsworth, N.R. (1995). Anti-diabetic Plants and Their Active Constituents. Phytomedicine. 2 (2): 137 189.
- 14. De Sales PM, De Souza PM, Simeoni LA, Magalhaes PO, Silveira D (2012) α-amylase inhibitors: A review of raw material and isolated compounds from plant source. J Pharm Pharm Science. 15: 141 183.
- 15. Karthic, K., Kirthiram, K.S., Sadasivam, S. and Thayumanavan, B. (2008). Identification of α- amylase inhibitors from Syzygium cumini Linn seeds. Indian J Exp Biology 46: 677 680.
- 16. Subramanian, R., Asmawi, A.Z. and Sadikun. A. (2008). In vitro alpha-qlucosidase and alpha- amylase enzyme

- inhibitory effects of Andrographis paniculata extract and andrographolide. J Pol Biochem Soc. 55: 391 398.
- 17. Playford, R.J., Pither, C., Gao, R. and Middleton, S.J. (2013). Use of the alpha glucosidase inhibitor acarbose in patients with 'Middleton syndrome': normal gastric anatomy but with accelerated gastric emptying causing postprandial reactive hypoglycemia and diarrhea. Can J Gastroenterol. 27 (7): 403 404.
- 18. Haslam, E. (1996). Natural polyphenols (vegetable tannins) as drugs: possible modes of action. Journal of Natural Products. 59: 205 215.
- 19. Jung, M., Park, M., Chul, H.L, Kang, Y., Seok-Kang, E. and Ki-Kim, S. (2006). Antidiabetic agents from medicinal plants. Curr. Med. Chem. 13: 1 16.
- 20. Reddy, N.V.L.S., Anarthe, S.J. and Raghavendra, N.M. (2010). Effect of thermal treatment on phenolic content. J Res. Biomed. Sci. 1 (2): 72 75.
- 21. Ramkumar, K.M., Thayumanavan, B., Palvannan, T. and Rajaguru, P. (2010). Inhibitory effect of Gymnema Montanum leaves on α -glucosidase and α -amylase activities and their relationship with polyphenolic content. Med. Chem. Res. 19: 948 961.
- 22. Choudhury, A., Maeda, K., Murayama, R. and DiMagno, E.P. (1996). Character of a wheat amylase inhibitor preparation and effects on fasting human pancreaticobiliary secretions and hormones. Gastroenterology. 111: 1313 1320.
- 23. Kavitha, S., Kamaraj, M. and Rajeshwari, S. (2012). In vitro alpha amylase and alpha glucosidase inhibition activity of crude ethanol extract of Cissus arnottiana. Asian Journal of Plant Science and Research. 2 (4): 550 553.
- 24. Boivin, M., Zinsmeister, A.R., Go, V.L.and DiMagno, E.P. (1987). Effect of a purified amylase inhibitor on carbohydrate metabolism after a mixed meal in healthy humans. Mayo Clin Proc. 62: 249 – 255.
- 25. Mohammed, A., Adelaiye, A.B., Bakari, A.G. and Mabrouk, M.A. (2009). Antidiabetic and some hematological effects of ethylacetate and n-butanol fractions of Ganoderma lucidum aqueous extract in alloxan-induced diabetic Wistar rats. International Journal of Medicine and Medicinal Sciences. 1 (12): 530 535.
- 26. Kumanan, R., Manimaran, S., Saleemulla, K., Dhanabal, S.P. and Nanjan, M.J. (2010). Screening of bark of Cinnamomum tamala (Lauraceae) by using α-amylase inhibition assay for antidiabetic activity. Int J Pharmaceut Biomed Res. 1 (2): 69 72.
- 27. Bhat, M., Zinjarde, S.S., Bhargava, S.Y., Kumar, A.R. and Joshi, B.N. (2011). Antidiabetic Indian Plants: A Good Source of Potent Amylase Inhibitors. Evi Based Complement Alternate Med. 2011: 810207.
- 28. Bray, G.A. and Greenway, F.L. (1999). Current and potential drugs for treatment of obesity. Endocr Rev. 20: 805 879.